

Mini Review

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Pharmacological Agents in the Clinic: Trial and Error

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ABSTRACT

Following an explosion of molecular and cellular research in the last 50 years, the study of specific molecular targets in the context of heart failure has generated much excitement. With the incidence of heart failure rising worldwide, new pharmacological agents are eagerly anticipated as a key tool with the potential for significant impact. In this regard, the results of the keenly awaited Does Cyclosporine Improve Clinical Outcome in ST Elevation Myocardial Infarction Patients (CIRCUS) trial have just been made available. Despite a substantial programme of supporting experimental and preclinical evidence however, the results have proved negative. This is of great disappointment to the clinical and scientific community. Here, this is discussed in the context of previous studies and future directions explored.

TARGETING MYOCARDIAL REPERFUSION INJURY TO MITIGATE HEART FAILURE

Molecular treatment regimens were first introduced into standard clinical practice in the 1980's but have not changed substantially since.¹ Myocardial infarction is one of the most widespread causes of secondary events, heart failure and death^{2,3} and patients with ST-segment Elevation Myocardial Infarction (STEMI) are a particularly high risk group. The extent of cell death in infarction i.e. infarct size, is suggested to be a major determinant in the likelihood of survival.⁴⁻⁶ Since myocardial reperfusion injury contributes greatly to the final infarct size, treatments that mitigate this have garnered significant interest.

The trial discussed herein focuses on the use of cyclosporine, a pharmacologic inhibitor of cyclophilin D that is a major component of the mitochondrial Permeability Transition Pore (mPTP). In ischemic tissue, Adenosine triphosphate (ATP) is depleted and cell death ensues. Upon reperfusion, further injury occurs due to generation of reactive oxygen species and calcium cation overload in mitochondria. This results in opening of the mPTP and mitochondria dependent cell death.⁷ Inhibition of cyclophilin D by cyclosporine and subsequently mPTP opening reduces the severity of myocardial reperfusion injury in animal models.⁸⁻¹¹ Cyclosporine is in fact already in use as an immunosuppressant in organ transplantation and autoimmune disorders, though associated with toxicity in chronic administration. Of further concern, it has also been shown to impede the compensatory hypertrophy of the remote noninfarcted myocardium by additionally inhibiting cyclophilin A and calcineurin.^{12,13} However, these studies were conducted with multiple doses of cyclosporine; a single, lower dose of cyclosporine is sufficient to reduce infarct size without adverse consequences on compensatory remodelling.¹⁴ Thus cardiac studies have used single, acute doses to minimise off-target effects while attempting to retain benefits.

CYCLOSPORINE PHASE II TRIALS

Initial results in a small phase II trial indicated a reduction in infarct size in STEMI patients when cyclosporine was administered immediately before reperfusion (primary Percutaneous Coronary Intervention; PCI), n=30 treated with cyclosporine, n=28 controls. The primary endpoint, infarct size, was assessed by measuring the cardiac injury biomarkers creatine kinase

and troponin I and by Magnetic Resonance Imaging (MRI) in 11 control and 16 cyclosporine treated patients. While creatine kinase levels were significantly reduced by cyclosporine treatment, troponin I was not. MRI showed a significant reduction in infarct size, but this did not translate into any improvement in left ventricular (LV) ejection fraction three months later.¹⁵ A follow-up report on the same patient cohort at 6 months, showed a persistent reduction in infarct size in the cyclosporine treated group, with a minor reduction in diastolic LV volume and a slight, though non-significant trend for reduced wall thickness. Similar to 3 months, cardiac function (ejection fraction) was not improved.¹⁴ However in another trial, in conjunction with thrombolytic therapy in STEMI patients cyclosporine addition did not improve any outcomes tested up to 6 months (n=50, n=51 controls), nor was infarct size affected though this was measured only by creatine kinase release.¹⁶

CYCLOSPORINE PHASE III TRIAL: CIRCUS

A phase III trial was commenced with larger numbers; 395 patients in the cyclosporine group and 396 controls, to assess whether clinical outcomes were improved at 1 year.^{17,18} The primary outcome was a combination of death, worsening of heart failure during the first admittance, rehospitalization and an increase of 15% or more in LV end diastolic volume. However, cyclosporine showed no benefit on these outcomes and cardiac function and remodelling (left ventricular ejection fraction and end-diastolic or systolic volumes) did not differ between treated and control groups.¹⁷ Unfortunately infarct size was not reported in this study, although area at risk was estimated using angiography and was not changed.¹⁷ Notably, in contrast to the phase II trial,¹⁵ creatine kinase levels (a measure of cardiac damage and indirectly, infarct size) were not improved by cyclosporine treatment.¹⁷

Why the failure from phase II to III?

Discrepancies in the outcomes from phase II and III might be explained by a number of factors. First, the former was rather preliminary and carried out with very small numbers which limits statistical power and reproducibility. Secondly, key parameters such as infarct size and LV wall thickness that were measured in phase II are not yet reported or available for phase III, thus similar endpoints are not being compared. Given the potential deleterious effects of cyclosporine on compensatory hypertrophy in the remote myocardium, rigorous assessment of remote wall thickness and LV remodelling is particularly essential. In the 6 month follow-up from phase II, a small though non-significant decrease in wall thickness in remote myocardium was seen,¹⁴ but was not reported at phase III with larger patient numbers.¹⁷ This must be addressed to confirm that a single dose of cyclosporine does not impair reparative remodelling. In addition, values for LV mass are not available for either study, and this is an important parameter in assessing remodelling rather than relying solely on end-diastolic volumes. However, the authors state that a sub-population of phase III patients were

selected to evaluate infarct size and LV remodelling in a more stringent manner¹⁸ and publication of the results of these will permit more direct comparison and dissection of the reasons for failure in moving to phase III.

The formulation of cyclosporine used is also given weight as a confounding factor; previously being Sandimmune (Novartis, which uses Cremophor EL, a polyoxyethylated castor oil as a vehicle) and currently, CicloMulsion (NeuroVive Pharmaceutical, where the vehicle is a lipid emulsion). However, in contrast to the preliminary positive study¹⁵ Sandimmune was shown to decrease maximal respiration in muscle, though was an effect of the vehicle control itself rather than cyclosporine.¹⁹ In addition, it was not found to be cardioprotective or to reduce infarct size in humans¹⁶ or pigs,²⁰ although it did reduce infarct size in rabbits.²¹ With regards to CicolMulsion used in the recently reported negative study, lipid emulsion itself reduces infarct size and improves functional recovery to a greater extent than cyclosporine through inhibition of cyclophilin D, although downstream signalling events indicate divergent mechanisms in cardioprotection.²² This would rather indicate that an improvement might be expected whether from cyclosporine, the vehicle itself or both, but this simply did not translate into the clinic. In addition, further confounding factors might also exist, for example, the effects of cyclosporine, cardioprotective or otherwise appear to be influenced by anaesthesia protocols.^{20,21,23}

CLINICAL TRIALS: LOST IN TRANSLATION

The disappointing results of this trial also call into question the translational abilities of current pre-clinical protocols. Such testing rarely includes prevalent comorbidities and risk factors or existing clinical treatments; without this, the interactions and outcomes in humans cannot be predicted. Consideration of these interacting factors could in fact reveal a sub-population of patients that might benefit greatly from this treatment. In addition, drug delivery methods and formulations that are clinically the most practical and likely to be used should be closely recapitulated in animal models. Further, many of the primary and secondary outcomes measured in trials do not form part of pre-clinical studies. For reasons of cost, survival up to one year for example might not be preferred but how can mouse to man possibly translate when different end-points are measured?

Clinical trial design itself is a crucial factor and must be reflected in preclinical animal models. Selection of inclusion and exclusion criteria, along with primary and secondary outcome measurements can significantly alter trial results and outcomes must be used that are sufficiently powered by the sample size. In addition, transparent and full reporting of results from animal studies are key to improving translation. The recently published Animal Research: Reporting of *In Vivo* Experiments (ARRIVE) guidelines for improving study design and reporting of animal studies are intended to do just this.²⁴

More substantial target validation and mechanistic information across multiple model systems more relevant to the human heart failure and clinical environment is also required. A carefully selected panel of heart failure mouse models that incorporate common co-morbidities along with commonly used clinical measurements and outcomes must be used to provide a more reliable basis for further drug development. In parallel, use of human stem cell-derived cardiomyocytes can address the concern of translatability from mouse to man. Although a drawback of this is cellular immaturity, efforts are underway to better recapitulate more mature human myocardium and a number of usable models now exist such as Engineered Heart Tissue (EHTs) that better model many of the key characteristics of the intact human heart.^{25,26}

A unified workflow prior to clinical trials with worldwide input and adherence would also provide a much improved and clear pathway from target identification to the patient. Indeed, such approaches now exist in the US and recommendations for generating similar in Europe have been recently proposed.^{27,28} Computational tools also provide an unprecedented resource that should be fully utilised.¹

Finally, an important point to consider is simply that targeting opening of the mPTP alone is not sufficient to prevent progression to heart failure in some settings, discussed below.

TARGETING CELL DEATH TO MITIGATE HEART FAILURE

It has been noted that the results of the recently completed Cyclosporine A in Reperfused Acute Myocardial Infarction (CYCLE) study using Sandimmune are expected imminently although the primary end point is ST-segment resolution rather than clinical outcomes.²⁹ These authors also propose that discovery of more specific inhibitors of mPTP opening might yield more promising results.²⁹ Perhaps however, the opposite might be required. In considering cell death, cardiomyocyte loss is a defining feature not only of infarction (acute, high levels of cell death) but of heart failure in general (chronic, low levels of cell death).³⁰ 'Apoptotic' cell death, measured by Deoxyribonucleic acid (DNA) fragmentation and caspase activity is prevalent in the failing heart³¹⁻³³ in addition to 'necrotic' cell death, inferred on the basis of reparative fibrosis and a diffuse smearing pattern of DNA, suggesting the contributions of multiple mechanisms for cell death in end-stage heart failure.³² Further, it has been suggested that chronic, low levels of cardiomyocyte death are equally, if not more so, deleterious to heart function than acute high levels of cell death associated with myocardial ischemia and reperfusion injury. Up to 50% of LV cardiomyocytes can be lost in ischemia before heart failure occurs, while only up to 20% of sporadic myocyte loss/dropout is required in the setting of pressure overload.³⁴ In addition, chronic low levels of sporadic cardiac muscle cell death are sufficient to cause heart failure in mice, and inhibition of caspase-dependent cell death in this model was protective.³⁵ Diverse gain- or loss-of-function mutations and pharmacological interventions in murine models

directly implicate cell-death signaling pathways as relevant therapeutic targets in reducing the inexorable progression to heart failure.³⁶

In conclusion, in the setting of acute infarction, a pharmacological agent that inhibits cell death, salvages jeopardized myocardium and reduces infarct size would be expected to prove highly effective. However, the outcome of the recent cyclosporine trial would indicate otherwise.¹⁷ It should be noted that cyclosporine only targets one aspect of cell death³⁷ and that heart failure incorporates multiple aspects.³⁶ A dual inhibitor or combined inhibitors targeting both mitochondria dependent and independent cell death might provide a more productive method than either alone to significantly impact on heart failure progression. Even if early, acute cell death is limited during reperfusion, subsequent chronic, low levels of cell death will still contribute significantly to ensuing heart failure and death. Inhibition of both acute and chronic, ongoing sporadic cell death should therefore be targeted.

REFERENCES

1. Fiedler LR. Cardiovascular research: past, present and future. *Heart Res Open J.* 2015; 2(2): 70-81. doi: [10.17140/HROJ-2-112](https://doi.org/10.17140/HROJ-2-112)
2. White HD, Chew DP. Acute myocardial infarction. *Lancet.* 2008; 372(9638): 570-584.
3. Heidenreich PA, Albert NM, Allen LA, et al. Forecasting the impact of heart failure in the United States: a policy statement from the American Heart Association. *Circ Heart Fail.* 2013; 6(3): 606-619. doi: [10.1161/HHF.0b013e318291329a](https://doi.org/10.1161/HHF.0b013e318291329a)
4. Miller TD, Christian TF, Hopfenspirger RN, Hodge DO, Gersh BJ, Gibbons RJ. Infarct size after acute myocardial infarction measured by quantitative tomographic 99mTc sestamibi imaging predicts subsequent mortality. *Circ.* 1995; 92(3): 334-341. doi: [10.1161/01.CIR.92.3.334](https://doi.org/10.1161/01.CIR.92.3.334)
5. Simoons ML, Serruys PW, van den Brand M, et al. Early thrombolysis in acute myocardial infarction: Limitation of infarct size and improved survival. *J Am Coll Cardiol.* 1986; 7(4): 717-728. doi: [10.1016/S0735-1097\(86\)80329-1](https://doi.org/10.1016/S0735-1097(86)80329-1)
6. Brener SJ, Maehara A, Dizon JM, et al. Relationship between myocardial reperfusion, infarct size, and mortality: The INFUSEAMI (intracoronary abciximab and aspiration thrombectomy in patients with large anterior myocardial Infarction) trial. *JACC: Cardiovasc Intervent.* 2013; 6(7): 718-724. doi: [10.1016/j.jcin.2013.03.013](https://doi.org/10.1016/j.jcin.2013.03.013)
7. Murphy E, Steenbergen C. Mechanisms underlying acute protection from cardiac ischemia-reperfusion injury. *Physiol Rev.* 2008; 88: 581-609. doi: [10.1152/physrev.00024.2007](https://doi.org/10.1152/physrev.00024.2007)

8. Baines CP, Kaiser RA, Purcell NH, et al. Loss of cyclophilin D reveals a critical role for mitochondrial permeability transition in cell death. *Nature*. 2005; 434: 658-662. doi: [10.1038/nature03434](https://doi.org/10.1038/nature03434)
9. Duchen MR, McGuinness O, Brown LA, Crompton M. On the involvement of a cyclosporin A sensitive mitochondrial pore in myocardial reperfusion injury. *Cardiovasc Res*. 1993; 27: 1790-1794. doi: [10.1093/cvr/27.10.1790](https://doi.org/10.1093/cvr/27.10.1790)
10. Ibáñez B, Heusch G, Ovize M, Van de Werf F. Evolving therapies for myocardial ischemia/reperfusion injury. *J Am Coll Cardiol*. 2015; 65: 1454-1471. doi: [10.1016/j.jacc.2015.02.032](https://doi.org/10.1016/j.jacc.2015.02.032)
11. Ong SB, Samangouei P, Kalkhoran SB, Hausenloy DJ. The mitochondrial permeability transition pore and its role in myocardial ischemia reperfusion injury. *J Mol Cell Cardiol*. 2015; 78: 23-34. doi: [10.1016/j.yjmcc.2014.11.005](https://doi.org/10.1016/j.yjmcc.2014.11.005)
12. Oie E, Bjornerheim R, Clausen OP, Attramadal H. Cyclosporin A inhibits cardiac hypertrophy and enhances cardiac dysfunction during postinfarction failure in rats. *Am J Physiol Heart Circ Physiol*. 2000; 278: 2115-2123.
13. Youn TJ, Piao H, Kwon JS, et al. Effects of the calcineurin dependent signaling pathway inhibition by cyclosporin A on early and late cardiac remodeling following myocardial infarction. *Eur J Heart Fail*. 2002; 4: 713-718. doi: [10.1016/S1388-9842\(02\)00120-4](https://doi.org/10.1016/S1388-9842(02)00120-4)
14. Mewton N, Croisille P, Gahide G, et al. Effect of cyclosporine on left ventricular remodeling after reperfused myocardial infarction. *J Am Coll Cardiol*. 2010; 55(12): 1200-1205. doi: [10.1016/j.jacc.2009.10.052](https://doi.org/10.1016/j.jacc.2009.10.052)
15. Piot C, Croisille P, Staat P, et al. Effect of cyclosporine on reperfusion injury in acute myocardial infarction. *N Engl J Med* 2008; 359: 473-481. doi: [10.1056/NEJMoa071142](https://doi.org/10.1056/NEJMoa071142)
16. Ghaffari, B. Kazemi, M. Toluey, et al. The effect of pre-thrombolytic cyclosporine-A injection on clinical outcome of acute anterior ST-elevation myocardial infarction. *Cardiovasc Ther*. 2013; 31(4): e34-e39. doi: [10.1111/1755-5922.12010](https://doi.org/10.1111/1755-5922.12010)
17. Cung TT, Morel O, Cayla G, et al. Cyclosporine before PCI in patients with acute myocardial infarction. *N Engl J Med*. 2015; 373(11): 1021-1031. doi: [10.1056/NEJMoa1505489](https://doi.org/10.1056/NEJMoa1505489)
18. Mewton N, Cung TT, Morel O, et al. (Rationale and design of the Cyclosporine to Improve clinical outcome in ST-elevation myocardial infarction patients (the CIRCUS trial). *Am Heart J*. 2015; 169: 758. doi: [10.1016/j.ahj.2015.02.020](https://doi.org/10.1016/j.ahj.2015.02.020)
19. Sanchez H, Zoll J, Bigard X, et al. Effect of cyclosporin A and its vehicle on cardiac and skeletal muscle mitochondria: relationship to efficacy of the respiratory chain. *Br J Pharmacol*. 2001; 133: 781-788. doi: [10.1038/sj.bjp.0704129](https://doi.org/10.1038/sj.bjp.0704129)
20. Karlsson LO, Zhou AX, Larsson E, et al. Cyclosporine does not reduce myocardial infarct size in a porcine ischemia-reperfusion model. *J Cardiovasc Pharmacol Ther*. 2010; 15: 182-189. doi: [10.1177/1074248410362074](https://doi.org/10.1177/1074248410362074)
21. Krolkowski JG, Bienengraeber M, Weihrauch D, Warltier DC, Kersten JR, Pagel PS. Inhibition of mitochondrial permeability transition enhances isoflurane-induced cardioprotection during early reperfusion: role of mitochondrial KATP channels. *Anesth Analg*. 2005; 101: 1590-1596. doi: [10.1213/01.ANE.0000181288.13549.28](https://doi.org/10.1213/01.ANE.0000181288.13549.28)
22. Li J, Iorga A, Sharma S, et al. Intralipid, a clinically safe compound, protects the heart against ischemia-reperfusion injury more efficiently than cyclosporine-A. *Anesthesiology*. 2012; 117: 836-846. doi: [10.1097/ALN.0b013e3182665e73](https://doi.org/10.1097/ALN.0b013e3182665e73)
23. Pagel PS, Krolkowski JG. Transient metabolic alkalosis during early reperfusion abolishes helium preconditioning against myocardial infarction: restoration of cardioprotection by cyclosporin A in rabbits. *Anesth Analg*. 2009; 108: 1076-1082. doi: [10.1213/ane.0b013e318193e934](https://doi.org/10.1213/ane.0b013e318193e934)
24. Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. *PLoS Biol*. 2010; 8(6): e1000412. doi: [10.1371/journal.pbio.1000412](https://doi.org/10.1371/journal.pbio.1000412)
25. Robertson C, Tran DD, George SC. Concise review: maturation phases of human pluripotent stem cell-derived cardiomyocytes. *Stem Cells*. 2013; 31(5): 829-837. doi: [10.1002/stem.1331](https://doi.org/10.1002/stem.1331)
26. Schwan J, Campbell SG. Prospects for in vitro myofilament maturation in stem cell-derived cardiac myocytes. *Biomark Insights*. 2015; 10(Suppl 1): 91-103. doi: [10.4137/BMI.S23912](https://doi.org/10.4137/BMI.S23912)
27. Lefér DJ, Bolli R. Development of an NIH consortium for preclinical assessment of cardioprotective therapies (CAESAR): a paradigm shift in studies of infarct size limitation. *J Cardiovasc Pharmacol Ther* 2011; 16: 332-339. doi: [10.1177/1074248411414155](https://doi.org/10.1177/1074248411414155)
28. Lecour S, Bøtker HE, Condorelli G, et al. ESC working group cellular biology of the heart: position paper: improving the preclinical assessment of novel cardioprotective therapies. *Cardiovasc Res*. 2014; 104(3): 399-411. doi: [10.1093/cvr/cvu225](https://doi.org/10.1093/cvr/cvu225)
29. Hausenloy DJ, Yellon DM. Targeting myocardial reperfusion injury-the search continues. *N Engl J Med*. 2015; 373(11): 1073-1075. doi: [10.1056/NEJMe1509718](https://doi.org/10.1056/NEJMe1509718)
30. Bing OHL. Hypothesis: apoptosis may be a mechanism

for the transition to heart failure with chronic pressure overload. *J Mol Cell Cardiol.* 1994; 26(8): 943-948. doi: [10.1006/jmcc.1994.1115](https://doi.org/10.1006/jmcc.1994.1115)

31. Narula J, Haider N, Virmani R, et al. Apoptosis in myocytes in end-stage heart failure. *N Engl J Med.* 1996; 335(16): 1182-1189. doi: [10.1056/NEJM199610173351603](https://doi.org/10.1056/NEJM199610173351603)

32. Olivetti G, Abbi R, Quaini F, et al. Apoptosis in the failing human heart. *N Engl J Med.* 1997; 336(16): 1131-1141. doi: [10.1056/NEJM199704173361603](https://doi.org/10.1056/NEJM199704173361603)

33. Kytö V, Saraste A, Saukko P, et al. Apoptotic cardiomyocyte death in fatal myocarditis. *Am J Cardiol.* 2004; 94(6): 746-750. doi: [10.1016/j.amjcard.2004.05.056](https://doi.org/10.1016/j.amjcard.2004.05.056)

34. Nadal-Ginard B, Kajstura J, Anversa P, Leri A. A matter of life and death: Cardiac myocyte apoptosis and regeneration. *J Clin Invest.* 2003; 111(10): 1457-1459. doi: [10.1172/JCI200318611](https://doi.org/10.1172/JCI200318611)

35. Wencker D, Chandra M, Nguyen K, et al. A mechanistic role for cardiac myocyte apoptosis in heart failure. *J Clin Invest.* 2003; 111(10): 1497-1504. doi: [10.1172/JCI200317664](https://doi.org/10.1172/JCI200317664)

36. Fiedler LR, Maifoshie E, Schneider MD. Mouse models of heart failure: cell signaling and cell survival. *Curr Top Dev Biol.* 2014; 109: 171-247. doi: [10.1016/B978-0-12-397920-9.00002-0](https://doi.org/10.1016/B978-0-12-397920-9.00002-0)

37. Nakagawa T, Shimizu S, Watanabe T, et al. Cyclophilin D-dependent mitochondrial permeability transition regulates some necrotic but not apoptotic cell death. *Nature.* 2005; 434: 652-658. doi: [10.1038/nature03317](https://doi.org/10.1038/nature03317)