

Review

Corresponding author*Xuesong He, DDS, PhD**

Assistant Adjunct Professor

School of Dentistry

University of California Los Angeles

10833 Le Conte Ave., Los Angeles

CA 90095, USA

E-mail: xhe@ucla.edu

Volume 2 : Issue 3

Article Ref. #: 1000DOJ2116

Article History**Received:** September 6th, 2015**Accepted:** September 29th, 2015**Published:** September 30th, 2015**Citation**

Wu T, Shi W, Loewy Z, He X. Manag-

ing denture biofilm related diseases.

Dent Open J. 2015; 2(3): 80-86.doi: [10.17140/DOJ-2-116](https://doi.org/10.17140/DOJ-2-116)

Managing Denture Biofilm Related Diseases

Tingxi Wu¹, Wen yuan Shi¹, Zvi Loewy² and Xuesong He^{1*}¹*School of Dentistry, University of California Los Angeles, 10833 Le Conte Ave., Los Angeles, CA 90095, USA*²*Department of Pharmaceutical and Biomedical Sciences, Touro College of Pharmacy, New York, 230 W 125th St., New York, NY 10027, USA***ABSTRACT**

The oral cavity harbors more than 700 microbial species and is one of the most complex ecosystems ever described. While the majority of these microbes are considered commensal, some of them are responsible for oral infectious diseases such as dental caries, periodontitis, halitosis and stomatitis. The advancement of modern science has greatly furthered our understanding of oral microbes and their roles in host health and disease. It has also led to the development of new tools for early detection, effective treatment, and prevention of oral microbial infections. This perspective provides a general understanding of oral microbiology, and its clinical relationship to oral infectious diseases, with a specific focus on denture-related microbial infections. The perspective also discusses the potential for developing innovative interventions for managing denture-related disease based on recent advances in our understanding of oral microbiology and denture-associated biofilms.

KEYWORDS: Stomatitis; Oral microbiota; *Candida albicans*.**ABBREVIATIONS:** SAP: Aspartyl proteinases; PL: Phospholipase; PDT: Photodynamic therapy; STAMPs: Specifically Targeted Antimicrobial Peptides.**INTRODUCTION**

The association between microbes and oral diseases had long been suspected. Dr. W. D. Miller is generally recognized as the father of modern dental microbial pathogenesis. His 1890 seminal book titled *Microorganisms of the Human Mouth*¹ makes the first connection between bacteria in dental plaque and tooth decay, and remains a foundation of current understanding of dental disease. For a long time, oral microbes had been indiscriminately regarded as pathogens. In fact, their removal from the oral cavity has become the main objective of dentists. Not until recently, did we realize that like microbes associated with other parts of the human body, most of the oral microorganisms are commensal and might have protective role in preventing the colonization of pathogens.^{2,3} More importantly, increasing evidence suggests that oral infectious diseases such as dental caries and periodontitis are often the result of the disturbed host homeostasis, and an imbalanced oral microbial ecology often leads to overgrowth of otherwise low abundant opportunistic pathogens.^{4,5}

Recent advances in molecular biological techniques are broadening our understanding of bacterial diversity and the societal community interactions which occur between species in the oral cavity.⁶ This has led to tremendous advances in our understanding of oral microbiology and its involvement in health and disease, including tooth decay, gum diseases, as well as the diseases associated with artificial dental apparatus introduced through modern dentistry.^{6,7}

Copyright

©2015 He X. This is an open access article distributed under the Creative Commons Attribution 4.0 International License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

ADVANCING UNDERSTANDING OF ORAL MICROBIOLOGY THROUGH MOLECULAR BIOLOGICAL APPROACHES

Our bodies are home to a multitude of microbial organisms that form distinct microflora inhabiting the gut, skin, vagina and oral cavity. These microbial communities have been of great interest to scientists in recent years due to their impact on host health and disease. Increasing lines of evidence indicate that these commensal microbiotas have important metabolic, trophic, and protective functions and greatly affect the host's physiology and pathology.^{8,9} For example, the importance of the gut flora in digesting unutilized substrates, training the immune system and protecting against epithelial cell injury is well appreciated,¹⁰⁻¹² and we are beginning to understand its potential role in systemic diseases, such as inflammatory bowel disease^{13,14} and obesity.¹⁵

Molecular biological tools have been critically important for identifying the diversity of these host-associated microbiotas, including the oral microbial community.¹⁶ Prior to the availability of such tools, determining the diversity of complex microbiological communities, such as those of the oral microbiota had been essentially dependent on the ability to culture and identify individual organisms. However, we then realized that only a small fraction of the organisms comprising these microbial communities has been isolated.⁶ In fact, accumulating lines of evidence suggested that there are extensive physical and metabolic interactions between different microbial species within the same community, which are essential for the growth and persistence of certain microbes¹⁷ and make them recalcitrant to cultivation. The power of molecular biological approaches, such as culture-independent 16S rRNA gene sequencing-based methods allows us to identify yet-uncultivable species and provides a more comprehensive and detailed inventory of human oral microbiota.⁶

The studies using culture-independent approaches have revealed the sheer magnitude of the diverse microbes, including yet-uncultivable species residing within the oral cavity.^{18,19} The human mouth is estimated to harbor more than 700 different bacterial species, comprising one of the most complex microbial flora.¹⁸ The diversity of microorganisms that inhabit the oral cavity includes bacteria, archaea, protozoa and fungi.^{20,21} An interesting perspective regarding diversity of the oral flora is the presence of Archaea as a constituent.²² Phylogenetically, Archaea is among the oldest known type of prokaryotes; it has previously been isolated from ocean bottom, and yet also appears to be a colonizer of the human oral cavity with yet-to-be-determined role in oral microbial ecology.²³ The diversity of the microbial flora reflects tremendous genetic information and immense bio-physiological potential that may have huge impact on host health and disease. If we consider that an average bacterial species has 2,000-6,000 genes, then an oral bacterial population of some 700 individual species represents a pool of over 1 million genes, 10 times more than human host genes. This provides the oral microbial environment with a huge quantity of informa-

tion related to unique metabolic pathways, the generation and secretion of various factors that can control and modify their ecological niche, and factors that may impact function of the human host.

THE STRUCTURE OF DENTAL AND DENTURE PLAQUE

Bacteria in the oral cavity often reside within biofilms, such as those that form dental and gingival plaque.²⁴ For edentulous and partially edentulous individuals who wear dentures, a denture-associated biofilm, or denture plaque, forms on the denture surface and could potentially serve as a reservoir of pathogenic microbes for infections.⁷

Dental and denture plaque are not simple matrices. They consist of a diverse collection of microbial species, and furthermore have a highly organized structure in which different species can occupy specific sites or niches within the biofilms.²⁵ During dental biofilm formation, bacteria that are early colonizers, such as Streptococci (i.e., *S. gordonii*, *S. oralis*, etc.) with specific adhesins can effectively bind to proteins deposited as a pellicle coat on the tooth surface. This is followed by the subsequent recruitment of intermediate and late colonizing species through cell-cell coadhesion *via* specific adhesin-receptor interactions.²⁵ These specific bacterial physical associations eventually generate a highly structured microbial community, which we recognize as dental plaque or biofilms.^{5,6,26} Furthermore, bacterial species within dental biofilm are often engaged in extensive signaling and metabolic interactions to ensure their survival within the microbial community.⁶

Dental biofilms of healthy subjects harbor a commensal oral microbial community with properties that limit the invasive potential of opportunistic pathogens.^{3,27} And likemost ecological communities, once established, dental biofilm generally has a stable and controlled population of different organisms and displays resilience to environmental disturbance.^{28,29} However, as will be discussed in the following section, the microbial composition of denture biofilm flora and their pathogenic potentials could differ significantly from that of healthy dental biofilm, thus contributing to the pathogenesis of denture-biofilm related diseases, such as stomatitis.

CONNECTIONS BETWEEN MICROBES AND ORAL DISEASES ASSOCIATED WITH DENTURE WEARING

Denture stomatitis is a common disorder in subjects wearing dentures, which are prostheses that provide important functional and esthetic improvements for edentulous and partially edentulous patients.^{30,31} The disorder is characterized as inflammation and erythema of the oral mucosal areas covered by the denture. The current view regarding the etiology of denture stomatitis is that it is a multifactorial infectious disease. It involves a number of associative factors, including denture-induced trauma, continual denture wearing and denture plaque harboring pathogenic microbes, such as *Candida*.³⁰ Among those

factors, the microbial biofilm formed on the denture surface plays a significant role in contributing to the disease pathogenic process. Whereas the normal commensal oral microbial community could prevent infection by interfering with the invasive potential of opportunistic pathogens, this is altered in the denture biofilm. Indeed, the microbial composition of the biofilm which forms on denture surfaces differs significantly from that observed in the oral cavity of healthy individuals.^{32,33} This could be due to the fact that denture wearing can alter normal oral physiology by affecting normal salivary flow that plays an important role in shaping the microbial community.³⁴ Meanwhile, the older segment of the population often has a comparable higher proportion of denture wearer.³⁵ These individuals are more likely to have systemic health conditions and could have already had imbalanced oral microbial ecosystem due to disturbance in host homeostasis. Furthermore, a denture provides a unique abiotic surface for microbial colonization, which often leads to the development of a denture biofilm with microbial composition and structure different from normal oral biofilm.^{7,32}

Compared to the normal oral and dental biofilms, denture biofilms are associated with a much higher occurrence of *Candida* yeasts, particularly *Candida albicans*.³⁶ *C. albicans* is a commensal fungal species commonly colonizing human mucosal surfaces. It co-exists with diverse oral microbial species and has long been adapted to the human host.³⁷ In healthy individuals, *C. albicans* is usually a minor component of their oral microfloras. However, under conditions of immune dysfunction or local predisposing factors such as poor oral hygiene or ill-fitted dentures, colonizing *C. albicans* can become an opportunistic pathogen. In these patients, *C. albicans* becomes more predominant and invasive, causing recurrent mucosal infections such as denture stomatitis.³⁸ The presence of *C. albicans* on denture and oral mucosal surfaces of denture wearers is positively associated with denture stomatitis.³⁹ The virulence factors of *C. albicans* have been well documented.⁴⁰ Among them, multiple host recognition biomolecules, such as Als1p and Hwp1p,^{41,42} as well as the secreted enzymes, including Aspartyl proteinases (SAP)⁴³ and Phospholipase (PL)⁴⁴ have been shown to play important role in determining *C. albicans*' pathogenicity. Meanwhile, its polymorphic growth patterns⁴⁰ as well as phenotypic switching⁴⁵ have also been implicated in contributing to its virulence. While *C. albicans* infection cannot be claimed as the single causal pathogen for inducing denture stomatitis, it has a strong associative presence when the disorder occurs, and its eradication from denture and mucosal surfaces is associated with reversal of the condition.^{46,47} Hence, it is generally accepted that *C. albicans* is a main opportunistic pathogen which is involved in the development and pathogenesis of denture stomatitis. Meanwhile, certain bacterial species, such as *Prevotellasp.*, *Veillonellasp.* and *Staphylococcus sp.* have been found to be enriched in denture biofilms,^{48,49} although their potential role in denture stomatitis pathogenesis remains to be determined. More importantly, increasing lines of evidence indicate the extensive Candida-bacterial interactions, which could impact their pathogenicity.³⁷ For

example, co-infection of *C. albicans* and *Staphylococcus aureus* has been shown to lead to increased mortality in animal model.⁵⁰ A better understanding of the physiology of Candida and bacteria co-existence and the inter-kingdom Candida-bacterial interactions would shed light on the impact of polymicrobial infection on the etiology of denture-related stomatitis.

TREATMENT OF DENTURE-RELATED INFECTIOUS DISEASE

Plaque formed on the denture surface often serves as a reservoir of opportunistic pathogens, including *C. albicans* for infections. In addition to maintaining good general oral hygiene, the most recommended approach to managing and preventing microbial-related disease associated with denture use is for patients to maintain a high level of denture hygiene by appropriate cleaning.^{51,52} Common approaches to denture cleaning utilized by patients include brushing with abrasive cleansers, such as toothpastes, and washing or soaking dentures using commercial chemical cleansers with antimicrobial compounds designed for this purpose. The latter is preferred, as brushing with abrasive cleansers has been shown to be less effective for removal of the biofilm, and furthermore, can roughen the denture surfaces and result in more rapid bacterial adherence and biofilm growth.^{7,53} However, as observed with other biofilms, a problematic issue associated with denture plaque is that it reduces the effectiveness of antimicrobial, including antifungal treatment.⁵⁴ The mechanisms by which biofilm environments enhance antimicrobial resistance are not fully understood. However, putative mechanisms likely include decreased ability of the antimicrobial agents to penetrate and diffuse within the biofilm matrix, protective functions conferred to the putatively susceptible bacteria due to slower growth rates and even changes in phenotype, and perhaps protective factors secreted by other microbes within the biofilm community which can degrade the applied antimicrobial agents.⁵⁵ In addition, the materials used in the manufacture of dentures can also affect adherence and colonization by microbes, including *C. albicans*, as well as impact the efficacy of antimicrobial treatment on the biofilm.⁵⁶

Soaking dentures in an appropriate commercial cleanser has been shown to be effective in removing attached microbes without increasing surface roughness.⁵⁷ Overnight denture removal is also important for controlling denture plaque, as it isolates the denture from salivary secretion that provides nutrients for microbial growth of denture biofilm. In addition to maintaining denture hygiene, various antimicrobials, including imidazole (clotrimazole, ketoconazole), triazole (fluconazole, itraconazole) and polyene (nystatin, amphotericin B) antifungals for treating Candida, and antibiotics for treating bacterial pathogens were also recommended for controlling denture-related mucosal infections.^{58,59}

More recently, a new antifungal therapeutic approach Photodynamic (PDT) therapy has been used to treat denture stomatitis.⁶⁰ PDT uses a photosensitizing agent and light of appro-

appropriate wavelength. The interaction between the photosensitizer and light in the presence of oxygen produces reactive species that induce cell damage and death.⁶¹ In a recent clinical trial, PDT was shown to be as effective as topical nystatin in the treatment of denture stomatitis.⁶² Since PDT can effectively kill *Candida* species, including strains resistant to conventional antifungal agents,⁶³ it has been regarded as a promising method for the treatment of dental stomatitis. Recurrence of stomatitis is frequently observed within short period of time after stopping antifungal treatment.⁶⁴ This is likely due to reinfection by residual pathogens that remain within plaque on dentures and are resistant to treatment. Meanwhile, many patients failed to respond to the usual treatment, largely due to the development of drug resistance of *Candida* species. For patients with systemic diseases, such as type 2 diabetes mellitus or being immunocompromised,⁶⁵ they often show less responsiveness to the treatment as well. When treating these patients, combined efforts including antifungal treatment and improving patients overall health status are critical in determining the outcome.⁶⁶

The knowledge we are gaining from molecular biological studies of dental and denture biofilms is contributing to the development of novel therapeutic tools.^{6,67} One approach is to build on our ability to identify specific pathogenic organisms that inhabit the biofilm, and develop therapeutics that specifically target these organisms. An example of this approach undertaken by our research group is the development of STAMPs (Specifically Targeted Antimicrobial Peptides).^{68,69} A typical STAMP consists of two functional moieties conjoined in a linear peptide sequence: a nonspecific antimicrobial peptide serves as the killing moiety, whereas a species-specific binding peptide provides specific binding to a selected pathogen and facilitates the targeted delivery of the attached antimicrobial peptide. The feasibility of this approach has been demonstrated by the development of C16G2, a STAMP specifically targeting *S. mutans*, the bacterium known to cause dental caries. C16G2 has been shown to remove *S. mutans* within *in vitro* multi-species biofilms with high efficacy and specificity,^{68,70} and is under further animal and human evaluations.⁷¹ The successful demonstration of this targeted approach could serve as proof-of-concept for applying this technology to the treatment of denture-related *Candida* infections.

CONCLUSIONS

The past decade has witnessed significant advances in our understanding of oral microbiota. We now better understand the structural and functional complexity of dental and denture plaque, and a strong connection between oral microbial ecology and host health and disease has been established. It is well known that the control of microbial pathogens, such as *C. albicans* on dentures and in the oral cavity is critical for the oral health of denture wearers. Continued efforts using modern scientific methods will help us develop more diagnostic tools and therapeutic interventions for the identification, treatment and prevention of denture infections. New and improved approaches will

be able to treat and control denture infections with less physical damage to denture surfaces by providing improved mechanisms for killing and removing microorganisms in the denture biofilm. We can envisage products that will have targeted killing of selective pathogens without affecting other commensal species within the same denture biofilm. Finally, we can also expect to see new products that will be able to enhance natural oral immunity, and provide cavity protection or control gingival disease in dentate individuals, and other inflammatory disorders in denture wearers.

CONFLICTS OF INTEREST

The authors declare that Wenyan Shi is an employee of C3 Jian, Inc. which has licensed technologies from UC Regents that could be indirectly related to this research project.

ACKNOWLEDGEMENT

This work was supported by grants from the National Institute of Health (NIH-1-R01-DE020102 and NIH-1-R01-DE023810-01) a grant from the Natural Sciences Foundation of China (30672322) and a grant from the International Science and Technology Cooperation Program of China (2011DFA30940).

REFERENCES

1. Miller WD. The micro-organisms of the human mouth. Biel, Switzerland: Graphische Anstalt Schuler AG; 1890.
2. He X, Hu W, Kaplan CW, et al. Adherence to *Streptococcus* facilitates *Fusobacterium nucleatum* integration into an oral microbial community. *Microb Ecol.* 2012; 63(3): 532-542. doi: [10.1007/s00248-011-9989-2](https://doi.org/10.1007/s00248-011-9989-2)
3. He X, Tian Y, Guo L, et al. *In vitro* communities derived from oral and gut microbial floras inhibit the growth of bacteria of foreign origins. *Microb Ecol.* 2010; 60(3): 665-676. doi: [10.1007/s00248-010-9711-9](https://doi.org/10.1007/s00248-010-9711-9)
4. Darveau RP. Periodontitis: a polymicrobial disruption of host homeostasis. *Nat Rev Microbiol.* 2010; 8(7): 481-490. doi: [10.1038/nrmicro2337](https://doi.org/10.1038/nrmicro2337)
5. Marsh PD. Microbial ecology of dental plaque and its significance in health and disease. *Adv Dent Res.* 1994; 8(2): 263-271. doi: [10.1177/08959374940080022001](https://doi.org/10.1177/08959374940080022001)
6. Kuramitsu HK, He X, Lux R, et al. Interspecies interactions within oral microbial communities. *Microbiol Mol Biol Rev.* 2007; 71(4): 653-670. doi: [10.1128/MMBR.00024-07](https://doi.org/10.1128/MMBR.00024-07)
7. von Fraunhofer JA, Loewy ZG. Factors involved in microbial colonization of oral prostheses. *Gen Dent.* 2009; 57(2): 136-143.
8. Human Microbiome Project Consortium. Structure, function

- and diversity of the healthy human microbiome. *Nature*. 2012; 486(7402): 207-214. doi: [10.1038/nature11234](https://doi.org/10.1038/nature11234)
9. Turnbaugh PJ, Ley RE, Hamady M, et al. The human microbiome project. *Nature*. 2007; 449(7164): 804-810.
10. Fujimura KE, Slusher NA, Cabana MD, Lynch SV. Role of the gut microbiota in defining human health. *Expert rev antiinfe*. 2010; 8: 435-454.
11. Guarner F, Malagelada JR. Gut flora in health and disease. *Lancet*. 2003; 361: 512-519. doi: [10.1016/S0140-6736\(03\)12489-0](https://doi.org/10.1016/S0140-6736(03)12489-0)
12. Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell*. 2012; 148: 1258-1270. doi: [10.1016/j.cell.2012.01.035](https://doi.org/10.1016/j.cell.2012.01.035)
13. Tamboli CP, Neut C, Desreumaux P, Colombel JF. Dysbiosis in inflammatory bowel disease. *Gut*. 2004; 53: 1-4.
14. Elinav E, Strowig T, Kau AL. NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. *Cell*. 2011; 145: 745-757. doi: [10.1016/j.cell.2011.04.022](https://doi.org/10.1016/j.cell.2011.04.022)
15. Cox AJ, West NP, Cripps AW. Obesity, inflammation, and the gut microbiota. *Lancet Diabetes Endocrinol*. 2015; 3: 207-215. doi: [10.1016/S2213-8587\(14\)70134-2](https://doi.org/10.1016/S2213-8587(14)70134-2)
16. Rajendhran J, Gunasekaran P. Microbial phylogeny and diversity: Small subunit ribosomal RNA sequence analysis and beyond. *Microbiol Res*. 2011; 166(2): 99-110.
17. He X, McLeanb JS, Edlund A, et al. Cultivation of a human-associated TM7 phylotype reveals a reduced genome and epibiotic parasitic lifestyle. *Proc Natl Acad Sci USA*. 2015; 112: 244-249. doi: [10.1073/pnas.1419038112](https://doi.org/10.1073/pnas.1419038112)
18. Paster BJ, Olsen I, Aas JA, et al. The breadth of bacterial diversity in the human periodontal pocket and other oral sites. *Periodontol 2000*. 2006; 42: 80-87. doi: [10.1111/j.1600-0757.2006.00174.x](https://doi.org/10.1111/j.1600-0757.2006.00174.x)
19. Aas JA, Paster BJ, Stokes LN, et al. Defining the normal bacterial flora of the oral cavity. *J Clin Microbiol*. 2005; 43(11): 5721-5732. doi: [10.1128/JCM.43.11.5721-5732.2005](https://doi.org/10.1128/JCM.43.11.5721-5732.2005)
20. Avila M, Ojcius DM, Yilmaz O. The oral microbiota: living with a permanent guest. *DNA Cell Biol*. 2009; 28(8): 405-411. doi: [10.1089/dna.2009.0874](https://doi.org/10.1089/dna.2009.0874)
21. Dewhirst FE, Chen T, Izard J, et al. The human oral microbiome. *J Bacteriol*. 2010; 192(19): 5002-5017. doi: [10.1128/JB.00542-10](https://doi.org/10.1128/JB.00542-10)
22. Horz HP, Conrads G. Methanogenic Archaea and oral infections-ways to unravel the black box. *J Oral Microbiol*. 2011; 3: doi: [10.3402/jom.v3i0.5940](https://doi.org/10.3402/jom.v3i0.5940)
23. Huynh HT, Pignoly M, Nkamga VD, et al. The repertoire of archaea cultivated from severe periodontitis. *PLoS One*. 2015; 10(4): e0121565. doi: [10.1371/journal.pone.0121565](https://doi.org/10.1371/journal.pone.0121565)
24. Kolenbrander PE, Palmer RJ, Periasamy S, et al. Oral multispecies biofilm development and the key role of cell-cell distance. *Nat Rev Microbiol*. 2010; 8(7): 471-480. doi: [10.1038/nrmicro2381](https://doi.org/10.1038/nrmicro2381)
25. Kolenbrander PE, Andersen RN, Kazmierzak K, et al. Spatial organization of oral bacteria in biofilms. *Methods Enzymol*. 1999; 310: 322-332. doi: [10.1016/S0076-6879\(99\)10026-0](https://doi.org/10.1016/S0076-6879(99)10026-0)
26. Kolenbrander PE, Palmer RJ, Rickard AH, et al. Bacterial interactions and successions during plaque development. *Periodontol 2000*. 2006; 42: 47-79. doi: [10.1111/j.1600-0757.2006.00187.x](https://doi.org/10.1111/j.1600-0757.2006.00187.x)
27. He X, Hu W, He J, et al. Community-based interference against integration of *Pseudomonas aeruginosa* into human salivary microbial biofilm. *Mol Oral Microbiol*. 2011; 26(6): 337-352. doi: [10.1111/j.2041-1014.2011.00622.x](https://doi.org/10.1111/j.2041-1014.2011.00622.x)
28. Rasiah IA, Wong L, Anderson SA, et al. Variation in bacterial DGGE patterns from human saliva: over time, between individuals and in corresponding dental plaque microcosms. *Arch Oral Biol*. 2005; 50(9): 779-787. doi: [10.1016/j.archoralbio.2005.02.001](https://doi.org/10.1016/j.archoralbio.2005.02.001)
29. Zaura E, Keijsers BJ, Huse SM, et al. Defining the healthy core microbiome of oral microbial communities. *BMC Microbiol*. 2009; 9: 259. doi: [10.1186/1471-2180-9-259](https://doi.org/10.1186/1471-2180-9-259)
30. Gendreau L, Loewy ZG. Epidemiology and etiology of denture stomatitis. *J Prosthodont*. 2011; 20(4): 251-260. doi: [10.1111/j.1532-849X.2011.00698.x](https://doi.org/10.1111/j.1532-849X.2011.00698.x)
31. Arendorf TM, Walker DM. Denture stomatitis: A review. *J Oral Rehabil*. 1987; 14(3): 217-227.
32. Mantzourani M, Gilbert SC, Fenlon M, et al. Non-oral bifidobacteria and the aciduric microbiota of the denture plaque biofilm. *Mol Oral Microbiol*. 2010; 25(3): 190-199. doi: [10.1111/j.2041-1014.2009.00565.x](https://doi.org/10.1111/j.2041-1014.2009.00565.x)
33. Murakami M, Nishi Y, Seto K, et al. Dry mouth and denture plaque microflora in complete denture and palatal obturator prosthesis wearers. *Gerodontology*. 2013; 32(3): 188-194. doi: [10.1111/ger.12073](https://doi.org/10.1111/ger.12073)
34. Edgar M, Dawes C, O'Mullane D. Saliva and oral health. 3rd ed. London, UK: British Dental Association; 2004.

35. Listl S. Denture wearing by individuals among the older segment of European populations. *Int J Prosthodont*. 2012; 25(1): 15-20.
36. Budtz-Jorgensen E. Ecology of Candida-associated Denture stomatitis. *Microbial Ecology in Health and Disease*. 2000; 12: 170-185.
37. Morales DK, Hogan DA. Candida albicans interactions with bacteria in the context of human health and disease. *PLoS Pathog*. 2010; 6(4): e1000886. doi: [10.1371/journal.ppat.1000886](https://doi.org/10.1371/journal.ppat.1000886)
38. Azie N, Neofytos D, Pfaller M, et al. The PATH (Prospective Antifungal Therapy) Alliance(R) registry and invasive fungal infections: update 2012. *Diagn Microbiol Infect Dis*. 2012; 73(4): 293-300. doi: [10.1016/j.diagmicrobio.2012.06.012](https://doi.org/10.1016/j.diagmicrobio.2012.06.012)
39. Mizugai H, Isogai E, Hirose K, et al. Effect of denture wearing on occurrence of Candida species in the oral cavity. *The Journal of Applied Research*. 2007; 7(3): 250-254.
40. Calderone RA, Fonzi WA. Virulence factors of Candida albicans. *Trends Microbiol*. 2001; 9(7): 327-335. doi: [10.1016/S0966-842X\(01\)02094-7](https://doi.org/10.1016/S0966-842X(01)02094-7)
41. Fu Y, Rieg G, Fonzi WA, et al. Expression of the Candida albicans gene ALS1 in Saccharomyces cerevisiae induces adherence to endothelial and epithelial cells. *Infect Immun*. 1998; 66(4): 1783-1786.
42. Staab JF, Ferrer CA, Sundstrom P, et al. Developmental expression of a tandemly repeated, proline-and glutamine-rich amino acid motif on hyphal surfaces on Candida albicans. *J Biol Chem*. 1996; 271(11): 6298-6305. doi: [10.1074/jbc.271.11.6298](https://doi.org/10.1074/jbc.271.11.6298)
43. Monod M, Hube B, Hess D, et al. Differential regulation of SAP8 and SAP9, which encode two new members of the secreted aspartic proteinase family in Candida albicans. *Microbiology*. 1998; 144(Pt 10): 2731-2737. doi: [10.1099/00221287-144-10-2731](https://doi.org/10.1099/00221287-144-10-2731)
44. Ghannoum MA. Potential role of phospholipases in virulence and fungal pathogenesis. *Clin Microbiol Rev*. 2000; 13(1): 122-143. doi: [10.1128/CMR.13.1.122-143.2000](https://doi.org/10.1128/CMR.13.1.122-143.2000)
45. Pomes R, Gil C, Nombela C. Genetic analysis of Candida albicans morphological mutants. *J Gen Microbiol*. 1985; 131(8): 2107-2113.
46. Budtz-Jorgensen E. Clinical aspects of Candida infection in denture wearers. *J Am Dent Assoc*. 1978; 96(3): 474-479.
47. Webb BC, Thomas CJ, Willcox MD, et al. Candida-associated denture stomatitis. Aetiology and management: A review. Part 1. Factors influencing distribution of Candida species in the oral cavity. *Aust Dent J*. 1998; 43(1): 45-50. doi: [10.1111/j.1834-7819.1998.tb00152.x](https://doi.org/10.1111/j.1834-7819.1998.tb00152.x)
48. Campos MS, Marchini L, Bernardes LA, et al. Biofilm microbial communities of denture stomatitis. *Oral Microbiol Immunol*. 2008; 23(5): 419-424. doi: [10.1111/j.1399-302X.2008.00445.x](https://doi.org/10.1111/j.1399-302X.2008.00445.x)
49. Jafari AA, Fallah-Tafti AA, Fattahi-bafghi A, et al. The comparison of predominant oral micro-flora in subjects with and without complete denture referred to Yazd dentistry department. *Journal of Community Health Research*. 2014; 3(3): 195-203.
50. Carlson E. Effect of strain of Staphylococcus aureus on synergism with Candida albicans resulting in mouse mortality and morbidity. *Infect Immun*. 1983; 42(1): 285-292.
51. Markovic D, Puskar T, Tesic D, et al. Denture cleaning techniques in the elderly affecting the occurrence of denture-induced stomatitis. *Med Pregl*. 1999; 52(1-2): 57-61.
52. Khasawneh S, al-Wahadni A. Control of denture plaque and mucosal inflammation in denture wearers. *J Ir Dent Assoc*. 2002; 48(4): 132-138.
53. Charman KM, Fernandez P, Loewy Z, et al. Attachment of Streptococcus oralis on acrylic substrates of varying roughness. *Lett Appl Microbiol*. 2009; 48(4): 472-477. doi: [10.1111/j.1472-765X.2008.02551.x](https://doi.org/10.1111/j.1472-765X.2008.02551.x)
54. Chandra J, Mukherjee PK, Leidich SD, et al. Antifungal resistance of candidal biofilms formed on denture acrylic in vitro. *J Dent Res*. 2001; 80(3): 903-908.
55. Gilbert P, Das J, Foley I. Biofilm susceptibility to antimicrobials. *Adv Dent Res*. 1997; 11(1): 160-167. doi: [10.1177/08959374970110010701](https://doi.org/10.1177/08959374970110010701)
56. Li L, Finnegan MB, Ozkan S, et al. In vitro study of biofilm formation and effectiveness of antimicrobial treatment on various dental material surfaces. *Mol Oral Microbiol*. 2010; 25(6): 384-390. doi: [10.1111/j.2041-1014.2010.00586.x](https://doi.org/10.1111/j.2041-1014.2010.00586.x)
57. Li L, Kim Y, Su Y. Effects of various methods for cleaning Candida albicans biofilm formed on denture acrylic resin in vitro. *Int J Oral Med*. 2010; 37: 157-160.
58. Lombardi T, Budtz-Jorgensen E. Treatment of denture-induced stomatitis: A review. *Eur J Prosthodont Restor Dent*. 1993; 2(1): 17-22.
59. Lalla RV, Dongari-Bagtzoglou A. Antifungal medications or disinfectants for denture stomatitis. *Evid Based Dent*. 2014; 15(2): 61-62. doi: [10.1038/sj.ebd.6401032](https://doi.org/10.1038/sj.ebd.6401032)
60. Mima EG, Pavarina AC, Silva MM, et al. Denture stomatitis treated with photodynamic therapy: Five cases. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2011; 112(5): 602-608.

doi: [10.1016/j.tripleo.2011.05.019](https://doi.org/10.1016/j.tripleo.2011.05.019)

61. Donnelly RF, McCarron PA, Tunney MM. Antifungal photodynamic therapy. *Microbiol Res.* 2008; 163(1): 1-12. doi: [10.1016/j.micres.2007.08.001](https://doi.org/10.1016/j.micres.2007.08.001)

62. Mima EG, Vergani CE, Machado AL, et al. Comparison of photodynamic therapy versus conventional antifungal therapy for the treatment of denture stomatitis: A randomized clinical trial. *Clin Microbiol Infect.* 2012; 18(10): E380-388.

63. Dovigo LN, Pavarina AC, Mima EG, et al. Fungicidal effect of photodynamic therapy against fluconazole-resistant *Candida albicans* and *Candida glabrata*. *Mycoses.* 2011; 54(2): 123-130. doi: [10.1111/j.1439-0507.2009.01769.x](https://doi.org/10.1111/j.1439-0507.2009.01769.x)

64. Cross LJ, Williams DW, Sweeney CP, et al. Evaluation of the recurrence of denture stomatitis and *Candida* colonization in a small group of patients who received itraconazole. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2004; 97(3): 351-358. doi: [10.1016/j.tripleo.2003.10.006](https://doi.org/10.1016/j.tripleo.2003.10.006)

65. Wilson J. The aetiology, diagnosis and management of denture stomatitis. *Br Dent J.* 1998; 185(8): 380-384.

66. Dorocka-Bobkowska B, Zozulinska-Ziolkiewicz D, Wierusz-Wysocka B, et al. *Candida*-associated denture stomatitis in type 2 diabetes mellitus. *Diabetes Res Clin Pract.* 2010; 90(1): 81-86. doi: [10.1016/j.diabres.2010.06.015](https://doi.org/10.1016/j.diabres.2010.06.015)

67. He X, Lux R, Kuramitsu HK, et al. Achieving probiotic effects *via* modulating oral microbial ecology. *Adv Dent Res.* 2009; 21(1): 53-56. doi: [10.1177/0895937409335626](https://doi.org/10.1177/0895937409335626)

68. Eckert R, He J, Yarbrough DK, et al. Targeted killing of *Streptococcus mutans* by a pheromone-guided “smart” antimicrobial peptide. *Antimicrob Agents Chemother.* 2006; 50(11): 3651-3657. doi: [10.1128/AAC.00622-06](https://doi.org/10.1128/AAC.00622-06)

69. Eckert R, Sullivan R, Shi W. Targeted antimicrobial treatment to re-establish a healthy microbial flora for long-term protection. *Adv Dent Res.* 2012; 24(2): 94-97. doi: [10.1177/0022034512453725](https://doi.org/10.1177/0022034512453725)

70. Guo L, McLean JS, Yang Y, et al. Precision-guided antimicrobial peptide as a targeted modulator of human microbial ecology. *Proc Natl Acad Sci USA.* 2015; 112(24): 7569-7574. doi: [10.1073/pnas.1506207112](https://doi.org/10.1073/pnas.1506207112)

71. Sullivan R, Santarpia P, Lavender S, et al. Clinical efficacy of a specifically targeted antimicrobial peptide mouth rinse: targeted elimination of *Streptococcus mutans* and prevention of demineralization. *Caries Res.* 2011; 45(5): 415-428. doi: [10.1159/000330510](https://doi.org/10.1159/000330510)