Original Research



Evaluation of the Anti-Obesity Effect of a Vegan Complex Supplement (Sinetrol, Green Tea, Bergamot and Chromium Yeast) in High-Fat Diet-Induced Obese Rats

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ABSTRACT

Background

Obesity is a chronic inflammatory response that not only leads to metabolic diseases such as diabetes, cardiovascular diseases, and cancers but also increases the risk of death, which needs to be addressed. Citrus and green tea, polyphenol-rich plants, hold the potential for improving metabolic diseases. Therefore, this study aims to investigate the effects of a vegan complex (VC) (Sinetrol, Green tea, Bergamot and Chromium yeast) supplementation on anti-obesity.

Methods

Obesity was induced in Wistar rats through a high-fat diet (HFD), and they were then randomly divided into 5 groups: 1 normal diet group and 4 high-fat diet groups (interventions with VC supplementation at 0, 207, 414, 828 mg/kg), with 12 rats in each group. After 4 weeks of supplementation, growth parameters, adipose tissue weight, hepatic lipid accumulation, and blood biochemical parameters were analyzed.

Results

After 4 weeks of VC supplementation, obese rats exhibited a significant reduction in final body weight, weight change, feed efficiency, visceral adipose tissue weight (epididymal, perirenal, mesenteric), and body fat percentage. In hepatic lipid content, VC supplementation significantly reduced liver weight, total cholesterol (TC), and triglyceride (TG) levels. In addition, all three doses of VC supplementation significantly reduced blood levels of TC, TG, low-density lipoprotein-cholesterol (LDL-C), and non-esterified fatty acid (NEFA), with the high dose also significantly lowering fasting blood glucose levels.

Conclusion

VC supplementation can effectively counteract high-fat diet-induced obesity by reducing hepatic and blood lipid accumulation, making it suitable for development as an anti-obesity dietary supplement.

Keywords

Sinetrol; Green tea; Citrus bergamia; Chromium yeast; Obesity; High-fat diet.

INTRODUCTION

Citrus plants are popular worldwide due to their pleasant flavor and rich content of bioactive compounds, including phenolic compounds, terpenoids, vitamin E, vitamin C, and minerals.¹ Sinetrol is a polyphenolic extract derived from citrus plants, which includes red-orange, sweet orange, bitter orange, grapefruit, and guarana. It is produced through physical methods such as cold pressing, extraction, and centrifugation.² Sinetrol is rich in anthocyanins and flavonoids, with strong antioxidant, anti-inflammatory, blood glucose stabilization, and anti-obesity effects.³ Bergamot (*Citrus bergamia* R.) is also a type of citrus plant that grows in southern Italy. It mainly contains flavonoids such as naringin, neohesperidin, and neoeriocitrin.⁴ Recent studies have shown that bergamot polyphenols exhibited antioxidant responses *in vitro* and *in vivo* and reduced glucose, cholesterol, serum triglycerides, systemic inflammation, and improved endothelial function.⁵ In addition to their health properties, which include anti-inflammation, anti-obesity, and anti-oxidant effects, citrus plants have also been found to confer neuroprotective effects in dementia models, including Alzheimer's disease (AD).⁶

Tea (Camellia sinensis (L.) O. Kuntze), originating from southern China with a history spanning over a millennium, is a

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popular non-alcoholic beverage. Presently, over 3 billion people worldwide use to consume tea.⁷ Based on its processing methods, tea can be categorized into white, yellow, green, oolong, black, and dark tea. Among them, green tea has the lowest volatile content ($20 \mu g/g$), in comparison, black tea has the highest ($710 \mu g/g$).⁸ Green tea, known for its potent antioxidant properties, is rich in catechins (flavon-3-ol) family, including (–)-epigallocatechin gallate (EGCG), (–)-epigallocatechin (EGC), (–)-epicatechin gallate (ECG), and (–)-epicatechin (EC). Besides, green tea contains abundant flavonoids, polyphenols, phenolic acids, amino acids, proteins, and lipids, making it crucial in promoting metabolism, anti-cancer effects, anti-inflammatory properties, anti-microbial activity, and cardiovascular protection.⁹

The prolonged intake of a diet with excessive caloric content leads to insulin over-secretion and the development of insulin resistant.¹⁰ Previous studies have indicated that chromium supplementation enhanced insulin sensitivity, promoting the metabolism of carbohydrates and fats, thereby benefiting the stabilization of blood sugar levels and the prevention of obesity.¹¹ Chromium is a trace element in the human diet, and approximately 1% to 2% of chromium can be absorbed from foods such as meat, cereal, nuts, grains, molasses, and brewer's yeast.¹² The chromium yeast, known for its non-toxic safety, simple processing, and low production costs, is commonly used in dietary supplements.¹³

Obesity is the root cause of metabolic disorders such as type 2 diabetes mellitus (T2DM), non-alcoholic fatty liver disease (NAFLD), and cardiovascular diseases (CVD). Moreover, the high oxidative stress resulting from obesity lead to immune dysregulation, increasing the risk of cancer and autoimmune diseases.¹⁴ Obesity is not only prevalent in adults. Over the past 30 years, the incidence of obesity in children and adolescents has also doubled and quadrupled, respectively.¹⁵ While it is worth mentioning that weight loss in obese individuals can reduce the risk of mortality and various diseases. A weight reduction of 13% in obese individuals has been shown to decrease the risk of T2DM (41%), sleep apnea (40%), hypertension (22%), dyslipidemia (19%), and asthma (18%).¹⁶ This underscores the importance and necessity of weight loss for overall health.

Numerous studies had suggested that supplementation with plant extracts was beneficial for improving obesity. The dietary intake of a blend of bioactive compounds, aimed at targeting various mechanisms or enhancing their efficacy through synergistic interactions, has emerged as an innovative strategy for preventing and treating obesity.^{17,18} Thus, this study aims to develop an antiobesity dietary supplement combining Citrus extract (Sinetrol and bergamot) with green tea extract and yeast chromium. A 4-week supplementation trial was conducted on obese rats induced by a high-fat diet (HFD), and the changes in body weight, fat weight, and liver fat accumulation were analyzed.

MATERIAL AND METHODS

Experimental Supplement

Co., Ltd. (500 mg per tablet). The formula contained Sinetrol extract (polyphenols from citrus and guarana), green tea extract, *Citrus bergamia* extract, and chromium yeast. The VC supplement was prepared as a suspension in 0.5% carboxymethylcellulose (CMC) solution at three concentrations of 20.7, 41.4, and 82.8 mg/mL, administered at a volume of 1 mL per 100 g of body weight. The control group rats were provided with an equivalent volume of CMC solution.

Animal and Study Design

Male SD rats were obtained from BioLASCO Co., Ltd. (Taipei, Taiwan). A total of 70 rats, aged 6 weeks, were housed at the Animal Facility of China Medical University. The study was approved by the IACUC of China Medical University (No. 2021-400). The animal facility maintained a temperature of 22±2 °C and a 12-hour light-dark cycle (lights on at 8 AM and off at 8 PM). All rats were freely allowed to access the experimental diet and sterilized water throughout the experiment. After one week of acclimatization, the rats were divided into a normal diet (ND) group and a HFD group with 12 and 58 rats, respectively. The ND group was fed a standard diet with a caloric content of 2.85 Kcal/g (Altromin 1320, Altromin Spezialfutter GmbH & Co. KG, ImSeelenkamp, Germany). The HFD group was fed an HFD, which had a caloric content of 5.24 Kcal/g and a fat content of 34.9% (D12492, Research Diet Inc., New Brunswick, NJ, USA). After five weeks of diet induction, 10 rats in the HFD group that were either underweight or overweight were removed. The remaining rats were then divided into four groups, each consisting of 12 rats. These groups were oral gavage daily with CMC (control) and VC supplement at 1X, 2X, and 4X doses (207, 414, or 828 mg/kg, respectively) once per day. After 4 weeks of treatment, the experimental rats were sacrificed under 4% isoflurane with fasting overnight before and blood was withdrawn from the abdominal aorta for biochemical analysis. Liver and adipose tissues were collected, rinsed, and weighed.

Plasma Biochemistry Analysis

The blood samples were centrifuged at 4700 rpm for 15 minutes to obtain plasma for biochemical analysis. The analysis of aspartate amino transferase (AST), alanine amino transferase (ALT), triglyceride (TG), uric acid, and creatinine was performed using a commercial assay kit (Roche, Rotkreuz, Switzerland) and measured with a serum biochemical analyzer (COBAS MIRA, NY, USA). Blood glucose levels were determined using the blood glucometer CareSens II (i-SENS, Inc., Wonju-si, Korea), which utilizes the glucose oxidase enzyme method for glucose measurement. Non-esterified fatty acid (NEFA) was measured using a commercial assay kit (Non-Esterified Fatty Acid, RANDOX, County Antrim, UK). The concentrations of total cholesterol (TC), low-density lipoprotein-cholesterol (LDL-C), and high-density lipoprotein-cholesterol (HDL-C) were determined using commercial assay kits (Fortress Diagnostics Limited, Antrim, UK). Blood sodium and potassium concentrations were measured using commercial enzyme-based assay kits (Fortress Diagnostics Limited, Antrim, UK).

Liver Lipid Concentration Determination

The vegan complex (VC) supplement was provided by HealthTake

Lipids were extracted from the liver using the method described by



Folch et al.¹⁹ For the lipid extraction, 0.1 grams of liver tissue were taken and homogenized with 2 mL of Folch solution (chloroform: methanol=2:1). The mixture was allowed to stand at room temperature for 60 minutes, followed by centrifugation at 5000 rpm for 5 minutes. The upper liquid layer was transferred to a clean 1.5 mL centrifuge tube, and 0.2 mL of 0.9% NaCl was added to the liquid and mixed well. At this point, the liquid appeared cloudy. After centrifuging at 2000 rpm for 5 minutes, it separated into two layers. The lower layer was retained and dried with nitrogen at 55 °C, and then dissolved by adding 100 μ L of a solvent (tert-butyl alcohol: triton X-100: methanol=2:1:1) and heating at 65 °C for 15 minutes. The concentrations of TC and TG in liver were determined using commercial assay kits (Roche, Rotkreuz, Switzerland).

Statistical Analysis

The experimental data were analyzed using a one-way analysis of variance (one-way ANOVA). Post hoc analysis was conducted using Duncan's multiple range test. And the Shapiro–Wilk test was used to test the normality of the data. Significant difference between groups was determined when p < 0.05.

RESULTS

Effect of VC Supplement on the Body Weight in HFD-induced Obese Rat

Figure 1 shows the body weight change during 4 weeks of VC supplementation. As results, the weight of rats fed with HFD was significantly higher than that fed with ND after 4 weeks of diet induction (week 0 of treatment period). After 1 week of treatment, the weight of the HFD+VC-4X group was significantly lower than the HFD control group. After 2 weeks of treatment, three VC treatment groups had significantly lower body weight than the HFD control group.

Effect of VC Supplement on the Growth Parameters in HFD-induced Obese Rat

The growth parameters of rats during the treatment period are detailed in Table 1.

There was no significant difference in induced body weight between the four HFD-fed groups. However, after 4 weeks of treat-





Growth Parameters	ND	HFD				
		Control	VC-IX	VC-2X	VC-4X	
Induced BW (g)	394.8±29.1	479.6±22.8ª	478.4±20.4ª	477.6±25.2ª	477.0±29.7ª	
Final BW(g)	464.8±40.5	619.3±31.2ª	578.2±29.4 ^{ab}	566.1±35.4 ^{ab}	562.0±44.6 ^{ab}	
BW change (g)	70.0±15.2	39.7± 3.2ª	99.8±16.8 ^{ab}	88.5±17.4 ^{ab}	85.0±23.9 ^{ab}	
Feed intake (g/rat/day)	30.5±3.2	21.7±1.4ª	20.8±1.3ª	20.5±1.6ª	20.0±1.5ª	
Energy intake (kcal/rat/day)	86.8±9.0	113.6±7.4ª	109.6±6.9ª	108.0±8.6ª	106.4±8.1ª	
Feed efficiency (%)	8.1±1.2	23.0±1.3ª	17.1±2.5 ^{ab}	15.4±2.3 ^{ab}	15.0±3.5 ^{ab}	

induced BW. Feed efficiency (%)=[Body weight change (g)/total feed intake (g)]×100%. BW, body weight.



ment, the final body weight and body weight change were significantly lower in the three VC supplementation groups compared to the HFD control group. The feed intake and energy intake also showed no significant differences among the four HFD-fed groups, but feed efficiency was significantly lower in the three VC supplementation groups, demonstrating a dose-dependent effect.

Effect of VC Supplement on the Weight of Adipose Tissues in HFD-induced Obese Rat

The results of the VC supplement on the weight of adipose tissues (A-D) and body weight ratio (E) are presented in Figure 2. After 4 weeks of treatment, the weight of epididymal adipose tissue, peri-



renal adipose tissue, mesenteric adipose tissue, total body fat, and body weight ratio all exhibited a significant decrease in the three VC supplementation groups when compared to the HFD control group.

Effect of VC Supplement on the Liver Lipid in HFD-induced Obese Rat

The livers of diet-induced rats were collected and weighed after sacrifice, and the lipid content of the liver was analyzed (Figure 3). The liver weight was significantly lower after 4 weeks of VC supplementation at three doses (1X, 2X, and 4X) compared to HFD-fed rats. In comparison to the HFD control group, the TC content in the liver significantly decreased in the three VC supplementation groups, and the TG content significantly decreased in the 2X and 4X VC supplementation groups after 4 weeks of treatment.

Effect of VC Supplement on Plasma Biochemical Parameters in HFD-induced Obese Rat

We analyzed the biochemical parameters related to liver function, kidney function, lipid profile, glucose, and electrolyte balance after 4 weeks of treatment. As shown in Table 2, the levels of AST, ALT, HDL-C, creatinine, uric acid, Na⁺ and K⁺ did not exhibit significant changes after three doses of VC supplementation (1X, 2X, and 4X) when compared with the HFD control group. However, the levels of TC, LDL-C, TG, and NEFA significantly decreased in the three





The reported values are the mean±SD (n=12). One-way ANOVA was performed for the data analysis. If there was a significant difference between the groups, Duncan's test was used to compare means between specific two groups. a Significantly different from the ND group, p<0.05. b Significantly different from the group treated with HFD only, p<0.05.

Serum Biochemical Parameters	ND -	HFD				
		Control	VC-IX	VC-2X	VC-4X	
AST (U/L)	120.3±18.8	123.1±16.0	120.3±22.8	127.8±12.8	113.0±15.8	
ALT (U/L)	39.7±6.9	45.3±16.2	37.0±9.5	36.5±10.0	37.5±14.0	
TC (mg/dL)	65.2±11.2	79.0±10.2ª	64.8±16.2 ^b	62.9±12.3⁵	66.4±16.2 ^b	
LDL-C (mg/dL)	18.5±4.2	26.7±4.3ª	19.3±7.8⁵	16.8±4.8 ^b	18.8±6.9 ^b	
HDL-C (mg/dL)	36.5±6.3	39.8±4.3	34.7±6.3	34.7±6.3	34.4±8.2	
TG (mg/dL)	34.3±3.7	46.0±4.2ª	36.9±7.0 ^b	36.9±6.0 ^b	35.0±8.3 ^b	
NEFA (mg/dL)	0.55±0.12	1.05±0.28ª	0.75±0.16 ^b	0.74±0.15⁵	0.66±0.16 ^b	
Glucose (mg/dL)	127.9±21.7℃	170.8±28.1ª	154.2±21.4ª	149.6±31.2	141.9±26.6	
Creatinine (mg/dL)	0.4±0.1	0.4±0.1	0.3±0.0	0.4±0.1	0.4±0.1	
Uric acid (mg/dL)	0.7±0.2	0.8±0.3	0.7±0.2	0.8±0.3	0.7±0.3	
Na+ (mEq/L)	142.3±1.9	140.7±2.6	140.8±3.4	141.2±3.1	141.5±2.0	
K⁺ (mEq/L)	4.4±0.2	4.4±0.3	4.4±0.5	4.5±0.3	4.4±0.2	
The reported values are the a significant difference betw °Significantly different from t p<0.05 AST Aspartate amir	mean±SD (n=12 een the groups, D the ND group, p<	?). One-way ANOV uncan's test was (0.05. ^b Significantly ^{T-} Alanine amino tr	A was performed f used to compare n different from the consferase TC Total	for the data analys neans between spe group treated wit	is. If there was cific two groups h HFD only,	

lipoprotein-cholesterol; HDL-C: High density lipoprotein-cholesterol; TG: Triglyceride; NEFA: Non-esterified fatty acid.

VC treatment groups. Additionally, blood glucose content was significantly lower after 4 weeks of 4X VC supplementation.

DISCUSSION

Obesity increases the risk of infectious and other chronic diseases, making it a highly prioritized issue. According to guidelines from the USA and the UK, a weight loss of 5% to 10% have a clinical health benefits.¹⁶ This study conducted an obesity animal model using a

HFD-induction for 5 weeks, followed by a 4-week administration of VC supplement (207, 414, or 828 mg/kg) to investigate its antiobesity effects. The results revealed that rats induced with a HFD showed a 33.2% increase in body weight and a 289% increase in total body fat at the end of the trial compared to the ND group. Administration of 1X, 2X, and 4X the VC supplement significantly reduced the body weight (6.6%, 8.6%, 9.2%, respectively) and total body fat (20.2%, 24.6%, 30.2%, respectively) in obese rats (Figure 1 and 2). Previous study indicated that supplementation with Sinetrol (100



and 300 mg/kg) in leptin-deficient mice (ob/ob) for 7 weeks significantly reduced final body weight and weight gain. The 300 mg/kg of Sinetrol also significantly decreased the weight of abdominal adipose tissue, visceral adipose tissue, and epididymal adipose tissue.²⁰ A Korean clinical trial also demonstrated that 12 weeks of Sinetrol polyphenol extract intake (900 mg/day) significantly reduced weight (3.28%), body fat (9.73%), waist circumference (5.71%), and hip circumference (4.71%) in obese subjects.²¹ Mice fed with an HFD containing 0.32% green tea extract exhibited significant reductions in both body weight and liver fat after 8 weeks, indicating a preventive effect against obesity.²² Rats administered with HFD combined 12% (v/v) juice of *Citrus bergamia* orally exhibited a significant decrease in the elevated body weight, visceral adipose tissue, as well as liver and heart weights induced by a high-fat diet.²³ These studies suggest that VC supplement might exhibit an anti-obesity effect, as evidenced in various obesity animal models and obese patients.

Excessive calorie intake results in the storage of energy in the form of fat within the body. An HFD, due to its elevated calorie content, is prone to inducing weight gain. Moreover, when the body exhibits a higher efficiency in utilizing food, the probability of obesity tends to increase.²⁴ In this study, the groups supplemented with VC supplement did not show alterations in feed intake and water consumption, yet demonstrated lower weight gain and higher feed efficiency (Table 1). Previous research has indicated that HFD-induced mice administered neohesperidin (50 mg/ kg) derived from bergamot exhibited significant reductions in body weight, adipose tissue weight, and serum inflammatory markers (tumor necrosis factor-alpha (TNF-α), monocyte chemoattractant protein-1 (MCP-1)) without feed intake change, demonstrating an effective inhibition of obesity.²⁵ HFD-induced obese mice supplemented with green tea extract for 9 weeks (300 mg/kg) significantly downregulated the expression of genes associated with lipid synthesis in white fat tissue (SREBP1c, FAS, ACC1, SCD1) while upregulating genes related to thermogenesis (PGC-1a, UCP1) and fatty acid catabolism (HSL, ATGL, MGL).²⁶ Another study revealed that supplementing guarana extract (1 g/kg) for 8 weeks in HFD-induced obese mice significantly reduced the body weight without altering feed intake. It also increased the content of mitochondrial functional proteins and the expression of genes related to thermogenesis (Sirt1, CREB1, AMPKa2, PGC1a, Nrf2, UCP1).27 These studies suggest that VC supplementation could mitigate obesity through the regulation of lipid metabolism and mitochondrial activity without affecting feed intake.

The liver is an organ responsible for fatty acid synthesis. Accumulation of excessive fats in the liver triggers inflammatory reactions and increase the risk of diseases such as hepatic lipid infiltration, non-alcoholic steatohepatitis (NASH), liver fibrosis, and liver cancer.²⁸ Citrus plants contain abundant polyphenols that reduce the concentration of pro-inflammatory cytokines (TNF- α , interleukin 6 (IL-6), MCP-1, cyclooxygenase-2 (COX-2)) in the liver that inhibited HFD-induced non-alcoholic fatty liver disease (NAFLD).²⁹ Previous studies have also shown that administering citrus extract (0.3% and 0.9% w/w of diet) to HFD-induced obese rats for 6 weeks decreased hepatic lipid, TG, and TC accumulation, and reduced hepatic acetyl-CoA carboxylase concentration.³⁰ Similarly, *Citrus bergamia*, rich in

polyphenols, reduced hepatic lipid accumulation, inflammatory cytokines (TNF-a, IL-6), and lipid oxidation concentration (malondialdehyde, MDA) in the livers of obese rats induced by a high sugarfat diet. Moreover, it increased the activity of antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT).³¹ A 12-week study administering green tea extract to HFD-induced mice also indicated that the abundant EGCG in green tea significantly reduced hepatic TC accumulation. This effect was achieved by suppressing the generation of lysophospholipids (LPLs), thereby reducing liver inflammation. As a result, the hepatic lipid metabolism of mice approximated that of mice fed an ND.32 In this study, rats administrated with VC supplement exhibited lower liver weight, hepatic TG, and hepatic TC levels (Figure 3). It suggests that VC supplementation not only possesses anti-obesity properties but also has a preventive effect against NAFLD, and the health benefits are associated with the abundant polyphenolic compounds present in the supplement.

HFD induction not only increases the weight of adipose tissue, leading to weight gain, but also causes abnormalities in lipid metabolism, increasing the levels of TC and TG in the blood.³³ Previous studies have shown that supplementing citrus extract to rats for 42 days (0.25-0.75% of the diet) significantly reduced HFD-induced weight gain and hepatic lipid accumulation, exhibiting anti-obesity effects. It also regulated blood lipids by promoting the expression of genes related to cholesterol degradation, leading to a decrease in serum TC and LDL-C levels.³⁴ In this study, rats supplemented with VC also exhibited lower-levels of TC, TG, LDL-C, and NEFA (Table 2). Furthermore, the 4X-VC supplement had a lowering effect on fasting plasma glucose (Table 2).

Previous study indicated that obese subjects, after supplementing with Sinetrol extract for 12 weeks (900 mg/day), experienced reductions in weight and body fat and showed a decrease in abnormal fasting blood glucose levels.²¹ Additionally, it increased the content of Apo A1 to promote lipid metabolism.³⁵ In mice with HFD-induced obesity, supplementation with green tea extract for 6 weeks (0.1-1% of HFD) demonstrated anti-obesity effects, inhibiting the elevation of blood TC, TG, LDL-C, and glucose levels caused by HFD, and reducing liver inflammation.³⁶ Mice subjected to a high-fat high-sugar diet with chromium yeast (32 mg/ kg) for a continuous 8 weeks exhibited a significant reduction in body weight and serum glucose levels, accompanied by an increase in hepatic glycogen accumulation, which improved insulin function and promoted energy metabolism.³⁷ These studies collectively indicate that the effects of the VC supplement on the regulation of blood metabolic indicators are associated with its rich content of citrus, green tea extracts and chromium yeast.

This study confirmed the anti-obesity effect of the VC supplement, providing a new formula for weight control. The VC supplement can mitigate HFD-induced obesity by reducing the accumulation of fat tissue and liver lipids, however, the gut and neuron modulation of VC in the body and its effects in humans remain unclear. Therefore, further research is needed to investigate these limitations, and the findings of this study can serve as a dosage reference for future clinical trials.



CONCLUSION

In conclusion, supplementation with the VC for 4 weeks after inducing obesity with an HFD resulted in a reduction in the final body weight, weight change, feed efficiency, adipose tissue weight, and hepatic lipid accumulation in obese rats. This demonstrates the anti-obesity efficacy of the VC supplement, with a dose-dependent effect. Furthermore, the VC supplement administration also improved abnormal blood metabolic indicators. From the above, it could be inferred that the VC supplement is suitable for development as an anti-obesity dietary supplement, with an effective dose of 207 mg/kg rat, equivalent to a daily intake of 2000 mg of VC supplement for a 60 kg adult.

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CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

REFERENCES

1. Saini RK, Ranjit A, Sharma K, et al. Bioactive compounds of citrus fruits: A review of composition and health benefits of carotenoids, flavonoids, limonoids, and terpenes. *Antioxidants (Basel)*. 2022; 11(2): 239. doi: 10.3390/antiox11020239

2. Dallas C, Gerbi A, Tenca G, Juchaux F, Bernard FX. Lipolytic effect of a polyphenolic citrus dry extract of red orange, grapefruit, orange (SINETROL) in human body fat adipocytes. Mechanism of action by inhibition of cAMP-phosphodiesterase (PDE). *Phytomedicine*. 2008; 15(10): 783-792. doi: 10.1016/j.phymed.2008.05.006

3. Dallas C, Gerbi A, Elbez Y, Caillard P, Zamaria N, Cloarec M. Clinical study to assess the efficacy and safety of a citrus polyphenolic extract of red orange, grapefruit, and orange (Sinetrol-XPur) on weight management and metabolic parameters in healthy overweight individuals. *Phytother Res.* 2014; 28(2): 212-218. doi: 10.1002/ptr.4981

4. Russo M, Arigò A, Calabrò ML, Farnetti S, Mondello L, Dugo P. Bergamot (Citrus bergamia Risso) as a source of nutraceuticals: Limonoids and flavonoids. *Journal of Functional Foods.* 2016; 20: 10-19. doi: 10.1016/j.jff.2015.10.005

5. Maiuolo J, Bosco F, Guarnieri L, et al. Protective role of an extract waste product from citrus bergamia in an in vitro model of neurodegeneration. *Plants (Basel)*. 2023; 12(11): 2126. doi: 10.3390/ plants12112126

6. Matsuzaki K, Nakajima A, Guo Y, Ohizumi Y. A narrative review of the effects of citrus peels and extracts on human brain health and metabolism. *Nutrients.* 2022; 14(9): 1847. doi: 10.3390/nu14091847

7. Zhao T, Li C, Wang S, Song X. Green tea (Camellia sinensis): A

review of its phytochemistry, pharmacology, and toxicology. *Molecules*. 2022; 27(12): 3909. doi: 10.3390/molecules27123909

8. Yin P, Kong Y-S, Liu P-P, et al. A critical review of key odorants in green tea: Identification and biochemical formation pathway. *Trends in Food Science & Technology*. 2022; 129: 221-232. doi: 10.1016/j.tifs.2022.09.013

9. Farhan M. Green tea catechins: Nature's way of preventing and treating cancer. *Int J Mol Sci.* 2022; 23(18): 10713. doi: 10.3390/ijms231810713

10. Tzenios N, Chahine M, Tazanios M. Obesity and endometrial cancer: The role insulin resistance and adipokines. *Special journal of the Medical Academy and other Life Sciences*. 2023; 1(2). doi: 10.58676/sjmas.v1i2.12

11. Molz P, Molz WA, Dallemole DR, et al. Potential ameliorative effects of chromium supplementation on glucose metabolism, obesity, and genomic stability in prediabetic rat model. *Biol Trace Elem Res.* 2021; 199(5): 1893-1899. doi: 10.1007/s12011-020-02299-1

12. Tsang C, Taghizadeh M, Aghabagheri E, Asemi Z, Jafarnejad S. A meta-analysis of the effect of chromium supplementation on anthropometric indices of subjects with overweight or obesity. *Clin Obes.* 2019; 9(4): e12313. doi: 10.1111/cob.12313

13. Xin X, Han M, Wu Y, et al. Dietary supplemental chromium yeast improved the antioxidant capacity, immunity and liver health in broilers under high stocking density. *Animals (Basel)*. 2022; 12(17): 2216. doi: 10.3390/ani12172216

14. Zhang X, Gao L, Meng H, Zhang A, Liang Y, Lu J. Obesity alters immunopathology in cancers and inflammatory diseases. *Obes Rev.* 2023; 24(12): e13638. doi: 10.1111/obr.13638

15. Nakadate K, Kawakami K, Yamazaki N. Anti-obesity and antiinflammatory synergistic effects of green tea catechins and citrus beta-cryptoxanthin ingestion in obese mice. *Int J Mol Sci.* 2023; 24(8): 7054. doi: 10.3390/ijms24087054

16. Haase CL, Lopes S, Olsen AH, Satylganova A, Schnecke V, McEwan P. Weight loss and risk reduction of obesity-related outcomes in 0.5 million people: Evidence from a UK primary care database. *Int J Obes (Lond).* 2021; 45(6): 1249-1258. doi: 10.1038/s41366-021-00788-4

17. Pan MH, Yang G, Li S, et al. Combination of citrus polymethoxyflavones, green tea polyphenols, and Lychee extracts suppresses obesity and hepatic steatosis in high-fat diet induced obese mice. *MolNutr Food Res.* 2017; 61(11). doi: 10.1002/mnfr.201601104

18. Most J, Warnke I, Boekschoten MV, et al. The effects of polyphenol supplementation on adipose tissue morphology and gene expression in overweight and obese humans. *Adipocyte*. 2018; 7(3): 190-196. doi: 10.1080/21623945.2018.1469942



19. Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. *J Biol Chem.* 1957; 226(1): 497-509.

20. Lee M, Kwon HO, Choi SG, Bae MH, Kim O-K. The effects of sinetrol-XPur on lipolysis of leptin-deficient obese mice. *Journal of the Korean Society of Food Science and Nutrition*. 2017; 46(3): 389-393. doi: 10.3746/jkfn.2017.46.3.389

21. Park SJ, Sharma A, Bae MH, et al. Efficacy and safety of sinetrol-XPur on weight and body fat reduction in overweight or obese adults: A 12-week, randomized, double-blind, parallel, placebocontrolled trial. *J Med Food*. 2020; 23(3): 335-342. doi: 10.1089/ jmf.2019.4649

22. Ushiroda C, Naito Y, Takagi T, et al. Mice fed with a high-fat diet (HFD) containing 0.32% green tea extract exhibited significant reductions in both body weight and liver fat after 8 weeks, indicating a preventive effect against obesity. *Journal of Clinical Biochemistry and Nutrition.* 2019; 65(1): 34-46.

23. De Leo M, Piragine E, Pirone A, et al. Protective effects of bergamot (Citrus bergamia Risso & Poiteau) juice in rats fed with high-fat diet. *Planta Med.* 2020; 86(3): 180-189. doi: 10.1055/a-1070-9325

24. Han X, Choi S-I, Men X, et al. Anti-obesity activities of standardized ecklonia stolonifera extract in 3T3-L1 preadipocytes and high-fat-diet-fed ICR mice. *Applied Sciences*. 2022; 12(10): 5115. doi: 10.3390/app12105115

25. Lu JF, Zhu MQ, Zhang H, et al. Neohesperidin attenuates obesity by altering the composition of the gut microbiota in high-fat diet-fed mice. *EASEB J.* 2020; 34(9): 12053-12071. doi: 10.1096/ fj.201903102RR

26. Li M, Xu J, Zhang Y, et al. Comparative analysis of fecal metabolite profiles in HFD-induced obese mice after oral administration of huangjinya green tea extract. *Food Chem Toxicol.* 2020; 145: 111744. doi: 10.1016/j.fct.2020.111744

27. Lima NDS, Teixeira L, Gambero A, Ribeiro ML. Guarana (Paullinia cupana) stimulates mitochondrial biogenesis in mice fed high-fat diet. *Nutrients.* 2018; 10(2): 165. doi: 10.3390/nu10020165

28. Ke Z, Zhao Z, Zhao Y, et al. PMFs-rich citrus extract prevents the development of non-alcoholic fatty liver disease in C57BL/6J mice induced by a high-fat diet. *Journal of Functional Foods*. 2018; 47:

28-39. doi: 10.1016/j.jff.2018.05.032

29. Hu M, Zhang L, Ruan Z, Han P, Yu Y. The regulatory effects of citrus peel powder on liver metabolites and gut flora in mice with non-alcoholic fatty liver disease (NAFLD). *Foods.* 2021; 10(12): 3022. doi: 10.3390/foods10123022

30. Huang C-H, Hsiao S-Y, Lin Y-H, Tsai G-J. Effects of fermented citrus peel on ameliorating obesity in rats fed with high-fat diet. *Molecules.* 2022; 27(24): 8966. doi: 10.3390/molecules27248966

31. Siqueira JS, Nakandakare-Maia ET, Vieira TA, et al. Effect of bergamot leaves (Citrus bergamia) in the crosstalk between adipose tissue and liver of diet-induced obese rats. *Livers.* 2023; 3(2): 258-270. doi: 10.3390/livers3020017

32. Nam M, Choi MS, Choi JY, et al. Effect of green tea on hepatic lipid metabolism in mice fed a high-fat diet. *J Nutr Biochem.* 2018; 51: 1-7. doi: 10.1016/j.jnutbio.2017.09.002

33. Bagabaldo PAA, Atienza LM, Castillo-Israel KAT, et al. 'Saba' banana (Musa acuminata x balbisiana BBB Group) peel pectin supplementation improves biomarkers of obesity and associated blood lipid disorders in obese hypercholesterolemic mice. *Curr Res Food Sci.* 2022; 5: 251-260. doi: 10.1016/j.crfs.2022.01.016

34. Feng K, Zhu X, Liu G, et al. Dietary citrus peel essential oil ameliorates hypercholesterolemia and hepatic steatosis by modulating lipid and cholesterol homeostasis. *Food & Function.* 2020; 11(8): 7217-7230.

35. Cases J, Romain C, Dallas C, Gerbi A, Rouanet JM. A 12-week randomized double-blind parallel pilot trial of Sinetrol XPur on body weight, abdominal fat, waist circumference, and muscle metabolism in overweight men. *Int J Food Sci Nutr.* 2015; 66(4): 471-477. doi: 10.3109/09637486.2015.1042847

36. Zhou J, Yu Y, Ding L, Xu P, Wang Y. Matcha green tea alleviates non-alcoholic fatty liver disease in high-fat diet-induced obese mice by regulating lipid metabolism and inflammatory responses. *Nutrients.* 2021; 13(6): 1950. doi: 10.3390/nu13061950

37. Wang MT, Guo WL, Yang ZY, et al. Intestinal microbiomics and liver metabolomics insights into the preventive effects of chromium (III)-enriched yeast on hyperlipidemia and hyperglycemia induced by high-fat and high-fructose diet. *Curr Res Food Sci.* 2022; 5: 1365-1378. doi: 10.1016/j.crfs.2022.08.015

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