

## Mini Review

### \*Corresponding author

Renata Machado Pinto, MSc, MD

Endocrinologist

Department of Biology

Replicon Research Center

Catholic University of Goiás

Rua 01, 690, Apto 100

Setor Oeste, Goiânia-GO

CEP 74115-040, Brazil

Tel. 55 62 92637102

E-mail: [drarenamachado@gmail.com](mailto:drarenamachado@gmail.com)

Volume 2 : Issue 4

Article Ref. #: 1000OROJ2119

### Article History

Received: March 16<sup>th</sup>, 2016

Accepted: March 21<sup>st</sup>, 2016

Published: March 24<sup>th</sup>, 2016

### Citation

Pinto RM, Cominetti C, da Cruz AD. Basic and Genetic Aspects of Food Intake Control and Obesity: Role of Dopamine Receptor D2 TAQIA Polymorphism. *Obes Res Open J.* 2016; 2(4): 119-127. doi: [10.17140/OROJ-2-119](https://doi.org/10.17140/OROJ-2-119)

### Copyright

©2016 Pinto RM. This is an open access article distributed under the Creative Commons Attribution 4.0 International License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

# Basic and Genetic Aspects of Food Intake Control and Obesity: Role of Dopamine Receptor D2 TaqIA Polymorphism

Renata Machado Pinto, MSc, MD<sup>1,2,3\*</sup>; Cristiane Cominetti, MSc, PhD<sup>3,6</sup>; Aparecido Divino da Cruz, PhD<sup>1,2,4,5</sup>

<sup>1</sup>Department of Biology, Replicon Research Center, Catholic University of Goiás, Goiânia, Goiás, Brazil

<sup>2</sup>Postgraduate Program in Genetics, Catholic University of Goiás, Goiânia, Goiás, Brazil

<sup>3</sup>Health Sciences Graduate Program, Federal University of Goiás, Goiânia, GO, Brazil

<sup>4</sup>Laboratory of Human Cytogenetics and Molecular Genetics, Secretary of State for Health of Goiás (LACEN/SESGO), Goiânia, GO, Brazil

<sup>5</sup>Biotechnology and Biodiversity Graduate Program, University of Brasília, Brasília, DF, Brazil

<sup>6</sup>Nutrition and Health Graduate Program, Federal University of Goiás, Goiânia, GO, Brazil

### ABSTRACT

Regulation of food intake, energy expenditure and store are strikingly linked to obesity. Homeostatic control of food intake, hunger and satiety involves adipose and gastrointestinal hormones, such as leptin, insulin and ghrelin, which eventually affect neuronal signaling in the hypothalamus arcuate nucleus. On the other hand, hedonic control of food intake relates to substances such as opioids, endocannabinoids, gamma-aminobutyric acid, serotonin and dopamine, which act on the motivation and reward mechanisms. Dopamine is a precursor of noradrenaline and adrenaline and modulates a number of physiological functions, such as appetite, depending on the brain area and the type of receptor stimulated. It has been established as the main neurotransmitter of the hypothalamic reward system. Beyond the homeostatic and hedonic energy balance control, genetic aspects are also tightly involved in obesity pathophysiology. In this context, some Single Nucleotide Polymorphism (SNP) has been linked to common obesity. Here, we highlight the role of the dopamine receptor D2 gene TaqAI polymorphism, which affects the D2 receptor availability and has been associated to obesity. Therefore, the aim of this mini review is to cover basic aspects of food intake, energy balance, dopamine-related aspects, including genetic ones, and the relation with obesity.

**KEYWORDS:** Dopamine; *DRD2* gene; Genetic polymorphism; Obesity; Nutrigenetics.

**ABBREVIATIONS:** CNS: Central Nervous System; ARC: Arcuate Nucleus; GI: Gastrointestinal; GRP: Gastrin-releasing peptide; CCK: Cholecystokinin; PYY: Peptide YY; GLP1: Glucagon-like peptide-1; ApoAIV: apolipoprotein AIV; CART: Cocaine and amphetamine-regulated transcript; POMC: proopiomelanocortin;  $\alpha$ -MSH: alpha-melanocyte-stimulating hormone; NPY: neuropeptide Y; AgRP: agouti-related protein; PC1: prohormone Convertase 1; GABA: gamma-aminobutyric acid; DA: dopamine; NAc: accumbens nucleus; LHA: Lateral Hypothalamic Area; VMH: Ventromedial hypothalamic nucleus; RDS: Reward Deficiency Syndrome; ADHD: Attention Deficit Hyperactivity Disorder; SNP: Single Nucleotide Polymorphism; *NPY*: neuropeptide Y gene; *FTO*: fat mass and obesity-associated gene; *PPAR*: peroxisome proliferator-activated receptor gene; *APOE*: apolipoprotein E gene; *APOA1*: apolipoprotein A1 gene; *PLIN*: perilipin gene; *UCP1*: uncoupling protein 1 gene; *UCP2*: uncoupling protein 2 gene; *INSR*: insulin receptor gene; *ADIPOQ*: adiponectin gene; *IL6*: interleukin-6 gene; *RETN*: resistin gene; GWLS: Genome-Wide Linkage Studies; GWAS: Genome-Wide Association Studies; GIANT: The Genetic Investigation of ANthropometric Traits Consortium; *ANKK1*: ankyrin gene; BMI: Body Mass Index.

**FOOD INTAKE REGULATION AND OBESITY: HOMEOSTATIC AND HEDONIC CONTROL**

Obesity is a multifactorial condition influenced by genetic, endocrine-metabolic, environmental and psychological factors. A delicate balance between three main biochemical and behavioral processes maintains body weight: food intake, energy expenditure control and energy storage control.<sup>1</sup>

Regulation of food intake by the Central Nervous System (CNS) depends on the interaction of a homeostatic component that aims the balance between energy and nutrients, and a hedonic component, which seeks food-associate pleasure (Figure 1).

Homeostatic control of intake depends on the hormonal peripheral signaling produced in response to changes in nutrient concentrations. Leptin and insulin are the main hormonal adiposity signals, and by reaching the CNS trigger mechanisms that promote inhibition of food intake and increased energy expenditure.<sup>2</sup> On the other hand, hunger and satiety sensations are communicated to the CNS by gastrointestinal hormones. During prolonged fasting, the stomach produces ghrelin that acts on the hypothalamus as an orexigenic signal. After food intake, ghrelin concentration falls, giving rise to the secretion of anorectic hormones, such as cholecystokinin (CCK), peptide YY (PYY) and glucagon-like peptide-1 (GLP1).<sup>3,4</sup>

The main targets of peripheral adiposity, hunger and satiety signaling are neurons in the hypothalamus arcuate nucleus (ARC). In this nucleus there are orexigenic neurons that produce neuropeptide Y (NPY) and agouti-related protein (AgRP), in addition to anorectic neurons producing cocaine and amphetamine-regulated transcript (CART) and proopiomelanocortin (POMC), precursor of alpha-melanocyte-stimulating hormone ( $\alpha$ -MSH) by the action of prohormone

convertase 1 (PC1).<sup>1,5</sup> The melanocortin 4 receptor (MC4R) plays an important role in the intricate hypothalamic appetite control. When leptin binds to its receptor on POMC neurons,  $\alpha$ -MSH binds to MC4R, which produces a satiety signal. On the other hand, binding of AgRP to MC4R promotes increased intake.<sup>2,6</sup> Leptin activates the POMC neurons and inhibits AgRP neurons.<sup>2,6,7</sup> A scheme on the interaction of homeostatic intake control is depicted in Figure 2.

In humans, nutrition has not only physiological, but also social and behavioral roles. Food hedonic value is influenced by taste and previous experiences.<sup>4</sup> Intake of highly palatable foods (high in sugar and fat) is able to “deregulate” appetite homeostatic control, perpetuating the stimulus to eat, which causes the intake to be primarily mediated by hedonic and not homeostatic needs.<sup>7</sup> For decisions on food search, initiation and termination of meal to be properly taken, it is necessary the right integration between hypothalamic signals and cortical centers where substances such as opioids, endocannabinoids, gamma-aminobutyric acid (GABA), serotonin and dopamine (DA) act on the mechanisms of motivation and reward.<sup>2</sup>

Endogenous opioids as  $\beta$ -endorphin, enkephalin and dinorphin activate receptors in the accubens nucleus (NAc) disinhibiting orexigenic neurons in the Lateral Hypothalamic Area (LHA). Endocannabinoids impair leptin signaling, and interact with dopaminergic and opioid systems through the activation of CB1 receptors that inhibit melanocortin pathway.<sup>2,7</sup> Only the role of eating facilitator through its action on NPY neurons and consequent blockage of POMC transmission was attributed to GABA.<sup>8</sup> It is now known that GABA released by AgRP neurons is necessary to maintain a minimum level of appetite and the normal regulation of energy balance.<sup>9</sup>

Serotonin promotes satiety by acting directly in the ARC neurons, activating POMC and inhibiting AgRP neurons.

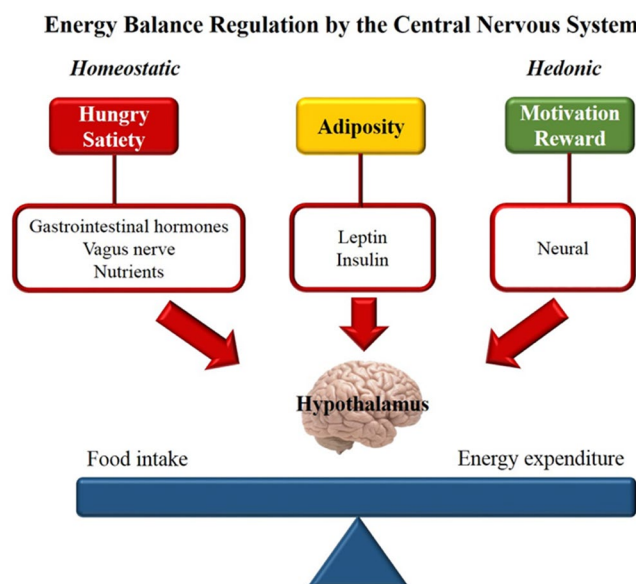
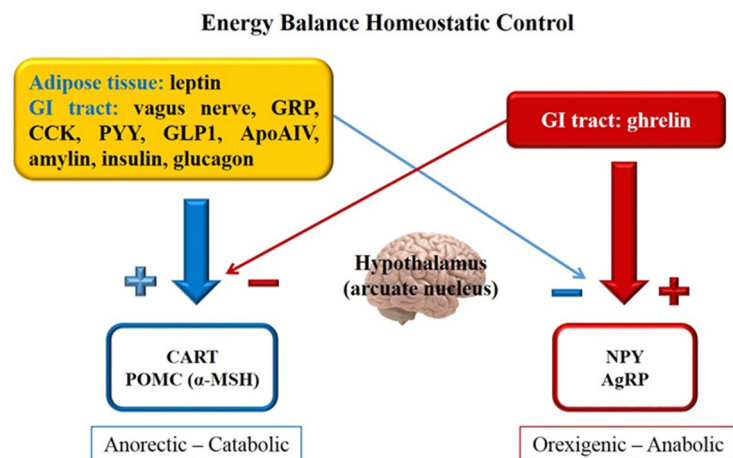


Figure 1: Homeostatic and hedonic control of energy balance.



GI, gastrointestinal; GRP, gastrin-releasing peptide; CCK, cholecystokinin; PYY, peptide YY; GLP1: glucagon-like peptide-1; ApoAIV, apolipoprotein AIV; CART, cocaine and amphetamine-regulated transcript; POMC, proopiomelanocortin; α-MSH, alpha-melanocyte-stimulating hormone; NPY: neuropeptide Y; AgRP, agouti-related protein.

Figure 2: Homeostatic regulation of energy balance.

It also inhibits orexins-producing neurons in LHA.<sup>2</sup>

DA is a catecholamine precursor of noradrenaline and adrenaline and is an endogenous neurotransmitter that modulates a number of physiological functions, including behavior, ion transport, vascular tone and blood pressure. Several experimental studies established DA as the main neurotransmitter of the reward system.<sup>10</sup> It is currently considered the “pleasure molecule” or the “anti-stress molecule.” Previc<sup>11</sup> established the concept of “dopaminergic society” and affirms that high DA concentrations was part of a general physiological adaptation to the increased meat consumption occurred two million years ago. The theory says that the “dopaminergic society” is characterized by high intelligence, personal destiny sense, religious/cosmic concerns and obsession with achieving goals.<sup>11</sup>

Regarding appetite, DA has varying effects depending on the brain area and the type of receptor stimulated. It has anorectic effect when it operates in the ARC, LHA and NAc, but acts as orexigenic in the ventromedial hypothalamic nucleus (VMH).<sup>2</sup> Several studies have related the dopaminergic brain circuits in eating behavior.<sup>7</sup> Animal studies reveal that the consumption of high-sugar or high-fat meals promotes the DA release in NAc.<sup>7,12</sup> The intake of a tasty meal for humans induces the DA release in a magnitude proportional to the meal degree of pleasure.<sup>13</sup>

**OBESITY AS PART OF THE “REWARD DEFICIENCY SYNDROME – RDS”**

Structures and cortico-limbic-striatal circuits form the brain reward system. Pleasurable stimuli activate this system and lead the individual to seek positive reinforcement of every type, not only food.<sup>14</sup> The brain reward cascade starts in the hypothalamus, where serotonin acts as a neurotransmitter stimulating the enkephalin release, which in turn inhibits GABAergic neurons in the substantia nigra. These GABAergic

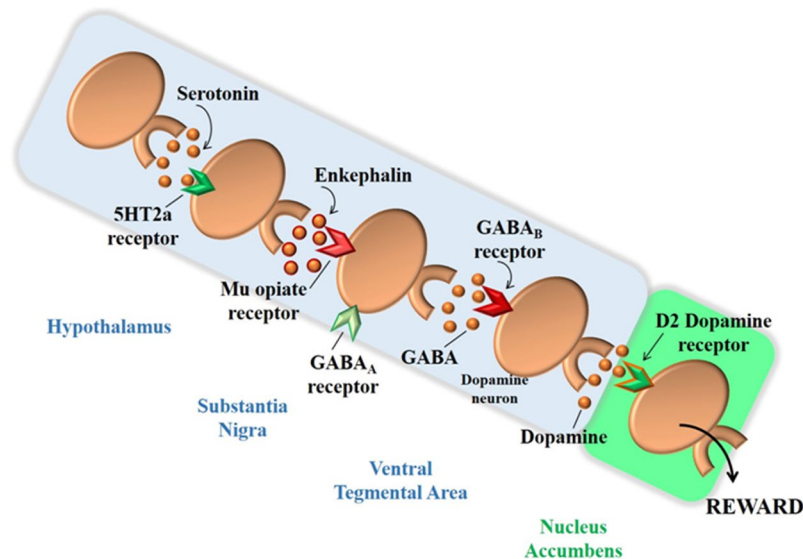
neurons act in the fine adjustment of DA amount that will be released in the NAc, the brain reward site (Figure 3).<sup>10</sup>

Studies show that low brain DA concentrations relate to greater vulnerability to substance abuse and abnormal behavior. It is known that all addictive drugs, as well as gambling, sex, food and even music promote DA release in the brain reward site.<sup>10</sup> In 1996, the term “Reward Deficiency Syndrome – RDS” was established, in order to define hypodopaminergic states-associated behaviors, which predispose to obsessive-compulsive behaviors and to impulsiveness.<sup>15</sup> The following changes are included in the RDS: a) Addictive behaviors: alcoholism, multiple substances abuse, obesity, smoking; b) Impulsive behaviors: attention deficit hyperactivity disorder (ADHD), Tourette’s syndrome, autism; c) Compulsive behaviors: abnormal sexual behavior, addiction to gambling and betting; d) Personality disorders: conduct disorder, antisocial personality, aggressive behavior and generalized anxiety.<sup>15</sup>

**GENETIC FACTORS RELATED TO OBESITY**

Common obesity, also named exogenous obesity, is a complex disease with multifactorial etiology. Pleiotropic genetic syndromes and monogenic diseases account for only 1% of obesity cases.<sup>16,17</sup> The most common forms of monogenic obesity occur due to mutations in genes related to hypothalamic control system of energy balance, as leptin-melanocortin system, which result in changes in the concentration and/or activity of hormones, receptors and enzymes, including leptin and its receptor, POMC, MCR4 and PC1.<sup>18</sup> In addition, there may be mutations in genes affecting the hypothalamus development and therefore, promoting obesity.<sup>6</sup> It is also important to note that obesity may be a central component of several pleiotropic syndromes, such as Alstrom, Albright, Pader-Willi, Bardet-Biedel, Fragile X, among other syndromes.<sup>17</sup>

In complex diseases such as common obesity, it is



**Figure 3:** Interaction of various neurotransmitters that forms the "Brain Reward Cascade".  
Adapted from Blum et al<sup>10</sup>

necessary that genetic factors are associated with a favorable environment for the phenotype emergence. The "thrifty genotype" hypothesis, described by Neel<sup>19</sup> proposes that genetic variations that result in higher capacity to store energy as fat were positively selected in food deprivation times. It is believed that over thousands of years this "thrifty genotype" has perpetuated and was essential in mankind evolution. This theory suggests that genes included in the "thrifty genotype" are responsible for the great ability to accumulate energy as fat, the ability to save energy at critical periods, the capacity to "turn off" non-essential metabolic pathways and to facilitate the intake of large amounts of food whenever they are available.<sup>20</sup> Currently, this same "thrifty genotype" has become disadvantageous, due to the easy access to energy-dense foods and to the low energy expenditure, which could explain the current obesity epidemic.

In 2007 Speakman published the "predation release" hypothesis, as an alternative to the "thrifty genotype" theory.<sup>21</sup> Based on anthropological and epidemiological evidence, genetic screening and experimental research, the theory suggests that the greater skill of lean individuals selected those best adapted to the search for food and to escape from predators; until fire was discovered in the Paleolithic period, and there was a significant increase in body weight over time. The theory attributes this increase in weight not only to the cooking capacity and better palatability of foods, but mainly to the fact that the fire was able to keep out the main predators, reducing energy expenditure. It also suggests that the initial genetic network responsible for low weight and high body performance has been suppressed and lost over the millennia.<sup>20,21</sup>

Common forms of obesity result from the interaction between variations in different genes and a favorable environment. Generally, many studies suggest a strong genetic

component in human obesity.<sup>22-27</sup> Studies report that in response to low calorie diets, some individuals lose weight more easily than others, and those carrying the same genotype respond in a similar manner when exposed to the same diet. Researches with monozygotic twins show that heredity accounts for 40 to 70% of inter-individual variation in cases of common obesity.<sup>28</sup> Differences between individuals and their predisposition to weight gain indicate that common variations in genomic DNA sequence, represented mainly by the Single Nucleotide Polymorphisms (SNP), may be responsible for the weight gain.<sup>27,29</sup> However, despite its great importance, the search for genes that raise the risk for obesity has not been easy.<sup>28,30</sup> It is still a challenge for the scientific community to separate the genetic component from the environmental one in the etiology of this disease. Individuals who are more susceptible to accumulating fat can carry risk variants in genes that influence appetite control (*NPY*, *POMC*, *MC4R*, etc.), cellular machinery regulation (*FTO*, *DRD2*, etc.), lipid metabolism and adipogenesis (*PPAR*, *APOE*, *APOA1*, *PLIN*, etc.), energy expenditure (*UCP1*, *UCP2*), insulin signaling (*INSR*, etc.) and inflammation (*ADIPOQ*, *IL6*, *RETN*, etc.).<sup>18,27</sup>

The polygenic nature of common obesity makes the discovery of risk genes and their variants a challenging task. Different approaches have been developed to elucidate the genetic component of obesity, such as GWLS (Genome-Wide Linkage Studies), that include co-segregation studies of certain chromosomal regions with a trait or disease<sup>30</sup>; analysis of candidate genes involved in plausible physiological pathways; and GWAS (Genome-Wide Association Studies), that track markers throughout the genome to identify associated polymorphisms.<sup>28</sup> Through a meta-analysis of 37 GWLS, Saunders and co-workers concluded that this is not an effective approach to identify genetic variants for common obesity, as



they did not locate any locus with conclusive evidence.<sup>31</sup>

Studies on candidate genes intended to identify the relation between one or more polymorphisms and a phenotype. In obesity, genes involved in the regulation of food intake, energy expenditure, lipid and glucose metabolism and adipose tissue development have been studied. In addition, genes described in monogenic forms of obesity have been investigated for a possible role in the common obesity genesis.<sup>6</sup> However, replication of most results has been somehow inconsistent, and so the findings of candidate gene studies remain obscure.<sup>32</sup>

In GWAS, unlike in the candidate gene approach, no assumption of the investigated gene function is made. These studies are based on the association of several markers, usually SNP and the approach is particularly useful in common complex diseases, such as obesity and diabetes.<sup>6</sup>

The latest update of “the human obesity gene map”, performed by Rankinen and co-workers associated obesity with 253 loci after analysis of 61 GWAS.<sup>33</sup> The number of associations between SNPs and obesity has 127 candidate genes described, and of these, 22 genes are supported by more than five studies. The map shows loci in all chromosomes, except for the Y.<sup>33</sup>

A more recent meta-analysis from GIANT (The Genetic Investigation of ANthropometric Traits) Consortium conducted in adults<sup>34</sup> established 32 loci of susceptibility to Body Mass Index (BMI), several of which were confirmed in French and German children with severe obesity.<sup>35</sup> In 2015, 97 BMI-associated loci were identified in a GWAS with 339,224 subjects (~95.0% of European and ~5.0% of non-European descent) from 125 studies (82 with GWAS and 43 with Metachip results). From those 97 loci, 56 were novel.<sup>36</sup>

Specifically for the childhood early-onset obesity, studies show that heredity is an important factor.<sup>37,38</sup> A large study of the childhood obesity genetics evaluated 5,530 cases and 8,318 controls through the analysis of 14 scientific papers, and the strong genetic influence in the childhood obesity development was definitely verified.<sup>39</sup> Three novel loci were

identified in a meta-analysis with 47,541 children from 33 studies (discovery and replication phases).<sup>40</sup>

**GENETIC VARIATIONS IN THE DOPAMINERGIC REWARD SYSTEM: DOPAMINE RECEPTOR D2 GENE (*DRD2*)**

Taking into account that DA plays a crucial role in the brain reward circuit and is involved in food behavior, the study of genetic polymorphisms that affect the availability and secretion of DA has been standing out.

The *DRD2* gene is located on chromosomal region 11q22-q23, with about 66,097 base pairs. This gene encodes the D2 subtype receptor, a transmembrane protein that couples to G proteins and inhibits the activity of adenylate cyclase. In an alternative splicing mechanism, the *DRD2* encodes two molecularly distinct protein isoforms – D2S and D2L – which are co-expressed, although the D2L production is favored. These two isoforms differ by the presence of 29 additional amino acids at D2L.<sup>41</sup> Both forms of D2 receptor have distinct physiological functions. The D2L acts mainly in postsynaptic regions while the D2S has a presynaptic self-receiving function.<sup>42</sup> This gene was included in “the human obesity gene map” supported by five candidate genes studies performed only with adults.<sup>33</sup>

***DRD2* TAQIA POLYMORPHISM**

The *DRD2* is highly polymorphic and there are already many SNPs cataloged and described (Figure 4). However, increasing attention has been given to C32806T SNP (rs1800497), characterized by the exchange of a cytosine for a thymine in a repetitive region in *ANKK1* (gene codifying ankyrin), located downstream of the *DRD2*.<sup>43</sup> This SNP is also known as TaqIA and appears to affect the D2 receptor availability. Variant allele A1 (T) is associated to a reduced metabolic rate of glucose in dopaminergic areas of the human brain, which indicates a low activity of dopaminergic neurons.<sup>44</sup>

Variations in the expression and activity of dopaminergic receptors and in DA release are related to overeating and obesity. Mice with reduced density of *DRD2* receptors in the striatum

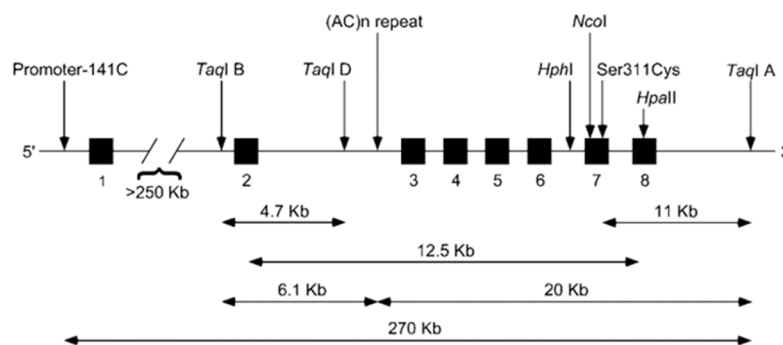


Figure 4: *DRD2* human gene with location of the most studied polymorphisms. Boxes represent exons and lines represent introns. Adapted from Noble, 2003.<sup>44</sup>

dorsal side, when fed with fat-rich diet gained more weight than those with normal density *DRD2*. The increase in *DRD2* mRNA expression in the nucleus accumbens and in the putamen ventral part of obese mice, when compared with obesity-resistant mice, seems to be a compensatory response to the *DRD2* pathway lower activation induced by overeating.<sup>45,46</sup>

Studies have suggested that obese individuals may have decreased DA availability through a mechanism of dopaminergic D2 receptors downregulation in the dorsal and lateral striatum.<sup>47-49</sup> Drugs blocking D2 receptors increase the appetite, and those that raise central AD concentration have anorectic effects.<sup>48</sup>

In addition, researches with adult humans suggest that increases in body mass are associated with the *DRD2* A1 allele.<sup>15,50,51</sup> In studies with positron emission tomography, the A1 allele was associated with lower density of the *DRD2*<sup>52</sup> and with reduction of glucose metabolism in human brain dopaminergic regions.<sup>53</sup> All RDS components, including obesity, were related to a low dopaminergic function due to the association with the *DRD2* A1 allele.<sup>10,15,54</sup>

The SNP C32806T in *DRD2* is also associated with a reduced dopaminergic brain activity<sup>55</sup> and the A1 allele was initially associated with BMI increase.<sup>15,56</sup> However, there are few studies<sup>28</sup> verifying the association of this polymorphism in children and adolescents.

It has been observed large variation in the allelic frequencies regarding the *DRD2* TaqIA SNP, even in populations of the same country. For example, in two studies with Turkish obese children the variant allele frequency was 51.0% in one<sup>57</sup> and only 20.0% in the other research.<sup>58</sup> In the Netherlands, the A1 allele frequency was 18.3% in obese children<sup>59</sup> and in North-American studies with obese children, it ranged from 17.0%<sup>60</sup> to 38.5%.<sup>61</sup> In a Brazilian study with obese and normal weight (controls) children, the A1 allele frequencies were 34.5% and 23.0%, respectively, and a statistically significant association between the A1 allele and obesity was verified.<sup>62</sup>

#### FINAL REMARKS

Food intake is controlled by an interrelation between homeostatic and hedonic factors. The search for food-associated pleasure involves the same neuronal pathway that is stimulated by addictive/compulsive behaviors (alcohol, gambling, sex and drugs), the so called “hypothalamic reward system”, that ends in DA. In this context, the minor A1 allele of the *DRD2* TaqIA polymorphisms has been associated with dopaminergic activity and increased obesity risk. Recognition of individuals predisposed to developing obesity through the determination of risk polymorphisms can guide prevention and treatment actions.

#### ACKNOWLEDGMENTS

We would like to thank Mr. Andre Hedlund for the English

review.

#### AUTHOR'S CONTRIBUTIONS

RMP, CC and ADC wrote and approved the final manuscript.

#### CONFLICTS OF INTEREST (COI) STATEMENT

The authors declare no conflicts of interest.

#### REFERENCES

1. Spiegelman BM, Flier JS. Obesity and the regulation of energy balance. *Cell*. 2001; 104(4): 531-543. doi: [10.1016/S0092-8674\(01\)00240-9](https://doi.org/10.1016/S0092-8674(01)00240-9)
2. Williams KW, Elmquist JK. From neuroanatomy to behavior: central integration of peripheral signals regulating feeding behavior. *Nat Neurosci*. 2012; 15(10): 1350-1355. doi: [10.1038/nn.3217](https://doi.org/10.1038/nn.3217)
3. Morton GJ, Cummings DE, Baskin DG, et al. Central nervous system control of food intake and body weight. *Nature*. 2006; 443(7109): 289-295. doi: [10.1038/nature05026](https://doi.org/10.1038/nature05026)
4. Velloso LA. The hypothalamic control of feeding and thermogenesis: implications on the development of obesity. *Arq Bras Endocrinol Metabol*. 2006; 50(2): 165-176. doi: [10.1590/S0004-27302006000200003](https://doi.org/10.1590/S0004-27302006000200003)
5. Schwartz MW, Woods SC, Porte DJ, et al. Central nervous system control of food intake. *Nature*. 2000; 404(6778): 661-671. doi: [10.1038/35007534](https://doi.org/10.1038/35007534)
6. Bougnères P. Genetics of obesity and type 2 diabetes: tracking pathogenic traits during the predisease period. *Diabetes*. 2002; 51(S3): S295-S303. doi: [10.2337/diabetes.51.2007.S295](https://doi.org/10.2337/diabetes.51.2007.S295)
7. Konturek SJ, Konturek PC, Konturek JW et al. Neuro-hormonal control of food intake; basic mechanisms and clinical implications. *J Physiol Pharmacol*. 2005; 56(S6): 5-25.
8. Damiani D, Damiani D. Sinalização cerebral do apetite. *Rev Bras Clin Med*. 2011; 9(2): 138-145.
9. Delgado TC. Glutamate and GABA in appetite regulation. *Front Endocrinol*. 2013; 4: 1-8. doi: [10.3389/fendo.2013.00103](https://doi.org/10.3389/fendo.2013.00103)
10. Blum K, Chen ALC, Giordano J, et al. The addictive brain: All roads lead to dopamine. *J Psychoactive Drugs*. 2012; 44(2): 134-143.
11. Previc F. *The Dopaminergic Mind in Human Evolution and History*. Oxford, Oxfordshire, UK: Cambridge University Press; 2009.
12. Avena NM, Rada P, Hoebel BG. Underweight rats have

- enhanced dopamine release and blunted acetylcholine response in the nucleus accumbens while bingeing on sucrose. *Neuroscience*. 2008; 156(4): 865-871. doi: [10.1016/j.neuroscience.2008.08.017](https://doi.org/10.1016/j.neuroscience.2008.08.017)
13. Small DM, Jones-Gotman M, Dagher A. Feeding induced dopamine release in dorsal striatum correlates with meal pleasantness ratings in healthy human volunteers. *Neuroimage*. 2003; 19(4): 1709-1715. doi: [10.1016/S1053-8119\(03\)00253-2](https://doi.org/10.1016/S1053-8119(03)00253-2)
14. King BM. The modern obesity epidemic, ancestral hunter-gatherers, and the sensory/reward control of food intake. *Am Psychol*. 2013; 68(2): 88-96. doi: [10.1037/a0030684](https://doi.org/10.1037/a0030684)
15. Blum K, Cull J, Braverman ER, et al. Reward deficiency syndrome. *American Scientist*. 1996; 84(2): 132-145.
16. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab*. 2004; 89(6): 2548-2556. doi: [10.1210/jc.2004-0395](https://doi.org/10.1210/jc.2004-0395)
17. Sabin MA, Werther GA, Kiess W. Genetics of obesity and overgrowth syndromes. *Best Pract Res Clin Endocrinol Metab*. 2011; 25(1): 207-220. doi: [10.1016/j.beem.2010.09.010](https://doi.org/10.1016/j.beem.2010.09.010)
18. Martinez JA. Perspectives on personalized nutrition for obesity. *J Nutrigenet Nutrigenomics*. 2014; 7(1): 1-3. doi: [10.1159/000365158](https://doi.org/10.1159/000365158)
19. Neel JV. Diabetes mellitus: a "thrifty" genotype rendered detrimental by "progress"? *Am J Hum Genet*. 1962; 14(4): 353-362.
20. Wang SP, Yang H, Wu JW et al. Metabolism as a tool for understanding human brain evolution: lipid energy metabolism as an example. *J Hum Evol*. 2014; 77: 41-49. doi: [10.1016/j.jhevol.2014.06.013](https://doi.org/10.1016/j.jhevol.2014.06.013)
21. Speakman JR. A Nonadaptive scenario explaining the genetic predisposition to obesity: the "predation release" hypothesis. *Cell Metab*. 2007; 6(1): 5-12. doi: [10.1016/j.cmet.2007.06.004](https://doi.org/10.1016/j.cmet.2007.06.004)
22. Bouchard C, Perusse L, Rice T, et al. The Genetics of Human Obesity. In: Bray GA, Bouchard C. *Handbook of Obesity*. New York, USA: Marcel Dekker; 1998: 157-190.
23. Rankinen T, Perusse L, Weisnagel SJ, et al. The human obesity gene map: the 2001 update. *Obes Res*. 2002; 10(3): 196-243. doi: [10.1038/oby.2002.30](https://doi.org/10.1038/oby.2002.30)
24. Hainer V, Zamrazilová H, Spálová J, et al. Role of hereditary factors in weight loss and its maintenance. *Physiol Res*. 2008; 57(Suppl. 1): S1-S15.
25. Day FR, Loos RJ. Developments in obesity genetics in the era of genome-wide association studies. *J Nutrigenet Nutrigenomics*. 2011; 4(4): 222-238. doi: [10.1159/000332158](https://doi.org/10.1159/000332158)
26. El-Sayed Moustafa JS, Froguel P. From obesity genetics to the future of personalized obesity therapy. *Nat Rev Endocrinol*. 2013; 9(7): 402-413. doi: [10.1038/nrendo.2013.57](https://doi.org/10.1038/nrendo.2013.57)
27. Walley AJ, Asher JE and FrCoguel P. The genetic contribution to non-syndromic human obesity. *Nat Rev Genet*. 2009; 10(7): 431-442. doi: [10.1038/nrg2594](https://doi.org/10.1038/nrg2594)
28. Böettcher Y, Korner A, Kovacs P, et al. Obesity genes: implication in childhood obesity. *Paediatr Child Health*. 2011; 22(1): 31-36. doi: [10.1016/j.paed.2011.08.009](https://doi.org/10.1016/j.paed.2011.08.009)
29. Bell CG, Walley AW, Froguel P. The genetics of human obesity. *Nat Rev Genet*. 2005; 6(3): 221-234. doi: [10.1038/nrg1556](https://doi.org/10.1038/nrg1556)
30. Loos RJ. Recent progress in genetics of common obesity. *Br J Clin Pharmacol*. 2009; 68(6): 811-829. doi: [10.1111/j.1365-2125.2009.03523.x](https://doi.org/10.1111/j.1365-2125.2009.03523.x)
31. Saunders CL, Chiodini BD, Sham P, et al. Meta-analysis of genome-wide linkage studies in bmi and obesity. *Obesity*. 2007; 15(9): 2263-2275. doi: [10.1038/oby.2007.269](https://doi.org/10.1038/oby.2007.269)
32. Shetty B, Shantaram M. Heritability of body weight: an evidence for obesity? *Int J Pharm Med Bio Sci*. 2014; 3(1): 15-20.
33. Rankinen T, Zuberi A, Chagnon YC, et al. The human obesity gene map: the 2005 update. *Obes Res*. 2006; 14(4): 529-644. doi: [10.1038/oby.2006.71](https://doi.org/10.1038/oby.2006.71)
34. Speliotes EK, Willer CJ, Berndt SI et al. Association analyses of 249,769 individuals reveal 18 new loci associated with body mass index. *Nat Genet*. 2010; 42(11): 937-948. doi: [10.1038/ng.686](https://doi.org/10.1038/ng.686)
35. Comuzzie AG, Cole AS, Laston SL, et al. Novel genetic loci identified for the pathophysiology of childhood obesity in the hispanic population. *Plos One*. 2012; 7(12): 1-9. doi: [10.1371/journal.pone.0051954](https://doi.org/10.1371/journal.pone.0051954)
36. Locke AE, Kahali B, Berndt SI, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature*. 2015; 518(7538): 197-206. doi: [10.1038/nature14177](https://doi.org/10.1038/nature14177)
37. Demerath EW, Choh AC, Czerwinski SA, et al. Genetics and environmental influences on infant weight and weight change: the fels longitudinal study. *Am J Hum Biol*. 2007; 19(5): 692-702. doi: [10.1002/ajhb.20660](https://doi.org/10.1002/ajhb.20660)
38. Griffiths LJ, Dezateux C, Cole TJ. Differential parental weight and height contributions to offspring birthweight and weight gain in infancy. *Int J Epidemiol*. 2007; 36(1): 104-107. doi: [10.1093/ije/dyl210](https://doi.org/10.1093/ije/dyl210)

39. Bradfield JP, Taal HR, Timpson NJ, et al. A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat Genet.* 2012; 44(5): 526-531. doi: [10.1038/ng.2247](https://doi.org/10.1038/ng.2247)
40. Felix JF, Bradfield JP, Monnereau C, et al. Genome-wide association analysis identifies three new susceptibility loci for childhood body mass index. *Hum Mol Genet.* 2016; 25(2): 389-403. doi: [10.1093/hmg/ddv472](https://doi.org/10.1093/hmg/ddv472)
41. Picetti R, Saiardi A, Abdel Samad T, et al. Dopamine D2 receptors in signal transduction and behavior. *Crit Rev Neurobiol.* 1997; 11(2-3): 121-142.
42. Usiello A, Baik JH, Rouge-Pont F, et al. Distinct functions of the two isoforms of dopamine D2 receptors. *Nature.* 2000; 408(6809): 199-203. doi: [10.1038/35041572](https://doi.org/10.1038/35041572)
43. Neville MJ, Johnstone EC, Walton, RT. Identification and characterization of ankk1: a novel kinase gene closely linked to DRD2 on chromosome band 11q23.1. *Hum Mutat.* 2004; 23(6): 540-545. doi: [10.1002/humu.20039](https://doi.org/10.1002/humu.20039)
44. Noble EP. D2 dopamine receptor gene in psychiatric and neurologic disorders and its phenotypes. *Am J Med Gene.* 2003; 116B(1): 103-125. doi: [10.1002/ajmg.b.10005](https://doi.org/10.1002/ajmg.b.10005)
45. Huang XF, Yu Y, Zavitsanou K, et al. Differential expression of dopamine d2 and d4 receptor and tyrosine hydroxylase mRNA in mice prone, or resistant, to chronic high-fat diet-induced obesity. *Brain Res Mol Brain Res.* 2005; 135(1-2): 150-161. doi: [10.1016/j.molbrainres.2004.12.013](https://doi.org/10.1016/j.molbrainres.2004.12.013)
46. Huang XF, Zavitsanou K, Huang X, et al. Dopamine transporter and D2 receptor binding densities in mice prone or resistant to chronic high fat diet-induced obesity. *Behav Brain Res.* 2006; 175(2): 415-419. doi: [10.1016/j.bbr.2006.08.034](https://doi.org/10.1016/j.bbr.2006.08.034)
47. Guo J, Simmons WK, Herscovitch P, Martin A, Hall KD. Striatal dopamine D<sub>2</sub>-like receptor correlation patterns with human obesity and opportunistic eating behavior. *Mol Psychiatry.* 2014; 19(10): 1078-1084. doi: [10.1016/j.bbr.2006.08.034](https://doi.org/10.1016/j.bbr.2006.08.034)
48. Wang GJ, Volkow ND, Logan J, et al. Brain dopamine and obesity. *Lancet.* 2001; 357(9253): 354-357. doi: [10.1016/S0140-6736\(00\)03643-6](https://doi.org/10.1016/S0140-6736(00)03643-6)
49. Hamdi A, Porter J, Chandan P. Decreased striatal D<sub>2</sub> dopamine receptors in obese Zucker rats: changes during aging. *Brain Res.* 1992; 589(2): 338-340. doi: [10.1016/0006-8993\(92\)91296-Q](https://doi.org/10.1016/0006-8993(92)91296-Q)
50. Nisoli E, Brunani A, Borgomainerio E, et al. D2 dopamine receptor (DRD2) gene Taq1A polymorphism and the eating-related psychological traits in eating disorders (anorexia nervosa and bulimia) and obesity. *Eat Weight Disord.* 2007; 12(2): 91-96.
51. Chen AL, Blum K, Chen TJ, et al. Correlation of the Taq1 dopamine D2 receptor gene and percent body fat in obese and screened control subjects: a preliminary report. *Food Funct.* 2012; 3(1): 40-48. doi: [10.1039/c1fo10089k](https://doi.org/10.1039/c1fo10089k)
52. Jönsson EG, Nöthen MM, Gründhage F, et al. Polymorphisms in the dopamine D2 receptor gene and their relationships to striatal dopamine receptor density of healthy volunteers. *Mol Psychiatry.* 1999; 4(3): 290-296.
53. Noble EP, Gottschalk LA, Fallon JH, et al. D<sub>2</sub> dopamine receptor polymorphism and brains regional glucose metabolism. *Am J Med Genet.* 1997; 74(2): 162-166. doi: [10.1002/\(SICI\)1096-8628\(19970418\)74:2<162::AID-AJMG9>3.0.CO;2-W](https://doi.org/10.1002/(SICI)1096-8628(19970418)74:2<162::AID-AJMG9>3.0.CO;2-W)
54. Blum K, Braverman ER, Wood RC, et al. Increased prevalence of the taq1 a1 allele of the dopamine receptor gene (DRD2) in obesity with comorbid substance use disorder: a preliminary report. *Pharmacogenetics.* 1996; 6(4): 297-305.
55. Noble EP, Blum K, Ritchie T, et al. Allelic association of the D2 dopamine receptor gene with receptor-binding characteristics in alcoholism. *Arch Gen Psychiatry.* 1991; 48(7): 648-654. doi: [10.1001/archpsyc.1991.01810310066012](https://doi.org/10.1001/archpsyc.1991.01810310066012)
56. Stice E, Yokum S, Bohon C, Marti N, Smolen A. Reward circuitry responsiveness to food predicts future increases in body mass: moderating effects of DRD2 and DRD4. *Neuroimage.* 2010; 50(4): 1618-1625. doi: [10.1016/j.neuroimage.2010.01.081](https://doi.org/10.1016/j.neuroimage.2010.01.081)
57. Ergun MA, Karaoguz MY, Koc A, et al. The apolipoprotein E gene and Taq1A polymorphisms in childhood obesity. *Genet Test Mol Biomarkers.* 2010; 14(3): 343-345. doi: [10.1089/gtmb.2010.0002](https://doi.org/10.1089/gtmb.2010.0002)
58. Araz NC, Nacak M, Balci SO, et al. Childhood obesity and the role of dopamine D2 receptor and cannabinoid receptor-1 gene polymorphisms. *Genet Test Mol Biomarkers.* 2012; 16(12): 1408-1412. doi: [10.1089/gtmb.2012.0244](https://doi.org/10.1089/gtmb.2012.0244)
59. Strien T, Snoek HM, van der Zwaluw C, Engels RC. Parental control and the dopamine D2 receptor gene (DRD2) interaction on emotional eating in adolescence. *Appetite.* 2010; 54(2): 255-261. doi: [10.1016/j.appet.2009.11.006](https://doi.org/10.1016/j.appet.2009.11.006)
60. Roth CL, Hinney A, Schur EA, et al. Association analyses for dopamine receptor gene polymorphisms and weight status in a longitudinal analysis in obese children before and after lifestyle intervention. *BMC Pediatr.* 2013; 13: 197-205. doi: [10.1186/1471-2431-13-197](https://doi.org/10.1186/1471-2431-13-197)
61. Duran-Gonzalez J, Ortiz I, Gonzales E, et al. Association study of candidate gene polymorphisms and obesity in a young Mexican-American population from south Texas. *Arch Med Res.* 2011; 42(6): 523-531.
62. Pinto RM, Silva DM, Queiroz FJ, et al. Reward deficiency



syndrome in children: obesity and metabolic disorders are associated with the SNP TaqIA C32806T of the DRD2 gene. *Obes Res Open J.* 2015; 2(2): 64-72. doi: [10.17140/OROJ-2-111](https://doi.org/10.17140/OROJ-2-111)